Gut microbial molecules in behavioural and neurodegenerative conditions

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Abstract | Mounting evidence suggests that the gut microbiome impacts brain development and function. Gut-brain connections may be mediated by an assortment of microbial molecules that are produced in the gastrointestinal tract, which can subsequently permeate many organs, including sometimes the brain. Studies in animal models have identified molecular cues propagated from intestinal bacteria to the brain that can affect neurological function and/or neurodevelopmental and neurodegenerative conditions. Herein, we describe bacterial metabolites with known or suspected neuromodulatory activity, define mechanisms of signalling pathways from the gut microbiota to the brain and discuss direct effects that gut bacterial molecules are likely exerting on specific brain cells. Many discoveries are recent, and the findings described in this Perspective are largely novel and yet to be extensively validated. However, expanding research into the dynamic molecular communications between gut microorganisms and the CNS continues to uncover critical and previously unappreciated clues in understanding the pathophysiology of behavioural, psychiatric and neurodegenerative diseases.

Gut microbiome-brain interactions

Decades passed before the principles of microbial pathogenesis took root after Louis Pasteur originally proposed a bacterial aetiology for infectious disease in the 1850s. Similarly, the pursuit of mechanistic evidence explaining the connections between intestinal bacteria and neurological disease has taken a century of research since hypothesized by Elie Metchnikoff and others in the early 1900s. The gut microbiota comprises bacteria and other microorganisms, including viruses, fungi, protists and archaea, that permanently or transiently inhabit the lower gastrointestinal tract, especially the small intestine and colon¹. The colon, in particular, is densely populated and teeming with dynamic metabolic activity, with a constant bidirectional flux of molecules between the microorganism and the host that extends beyond the gut into the entire body^{2,3}. This chemical 'factory' can affect the maternal environment during pregnancy and prenatally exposes the fetus to signals of microbial influence⁴. After birth, the gut microbiota is quickly established and

the community then stabilizes within the first 2 years of life¹, leading to lifelong and intimate crosstalk between the host and their microbial co-inhabitants. The level of diversity and the specific members of the microbiota can differ greatly between individuals and can shift within an individual depending on age, genetics, health status, diet and lifestyle⁵.

The gastrointestinal tract contains many diverse cell types in close proximity and is exposed on the luminal side to an external environment containing the dietary components and the gut microbiota. Within the gut tissue exist about 70% of the body's immune cells constantly sampling microbial components and maintaining homeostasis6, along with dense innervation along the gut by neurons that are housed entirely within intestinal tissue (108 intrinsic neurons7) as well as neurons connecting the gut to the spinal cord and brain. The vagus nerve, a principle neuronal connection between the gut and the brain, comprises a bundle of neurons that send and receive signals directly between gut tissue (and other organs) and the brainstem. These signals

can then be further transmitted throughout the brain.

Evidence that the gut microbiota influences brain development and function began to emerge with studies comparing conventionally colonized mice (also called specific pathogen-free mice) against mice in drastically altered microbial states, such as the complete absence of microbial exposure (germ-free mice). Additional insights were gained by using controlled introduction of a certain microorganism or community (gnotobiotic mice), or by treating conventional mice with broad-spectrum antibiotic cocktails that depleted their microbiome. Germ-free and antibiotic-treated animals exhibit altered levels of neurotrophic factors such as brain-derived neurotropic factor (BDNF) as well as abnormal neuropeptide and neurotransmitter levels⁸⁻¹³, all of which can, in turn, affect crucial neurodevelopmental processes such as neurogenesis, synaptogenesis and synaptic maturation and pruning, and neural activity^{8,9,14-17}. Gross morphology and volume of the brain also differs between specific pathogen-free and germ-free mice, especially in the amygdala, hippocampus and thalamus regions^{18,19}, with morphological changes observed at the cellular level in various cell types, including neurons, oligodendrocytes and microglia, in both germ-free and antibiotic conditions^{12,18,20,21}. Microbial exposure also alters the host neurological status and leads to changes in signalling pathways. For instance, the hypothalamic-pituitaryadrenal axis is dysregulated in germ-free and antibiotic-treated mice14, which results in an exaggerated glucocorticoid response. These hypothalamic-pituitary-adrenal axis changes are associated with some behavioural patterns in testing paradigms that model social activity²²⁻²⁴, anxiety^{9,25,26}, cognitive function and depressive^{25,27} behaviours^{10,11,24,28-30}

Gut microbial communities differ between individuals with certain health issues and healthy controls³¹. An imbalance in the gut microbial community is associated with various neurological and psychological disorders, although establishing which of these associations are causal relationships remains under investigation^{32–38} (FIG. 1). Recent work has shown that an altered

Neurological disorders with associated gut dysbiosis

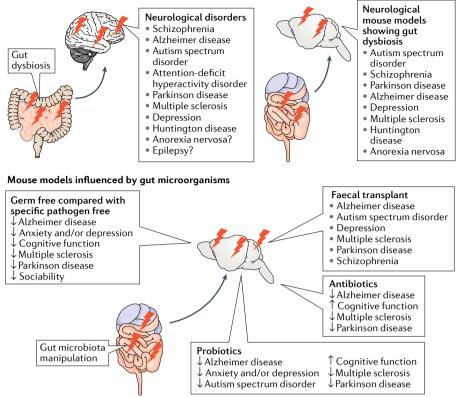


Fig. 1 | **Neurological disorders and models associated with shifts in gut microbiota.** Top: human conditions and mouse models with known differences in the gut microbial community compared with healthy controls. Question marks show disorders with implicated but less established changes to the microbiota. Bottom: rodent models shown to be improved, exacerbated or caused by manipulation of gut microorganisms. These are divided into four categories of experimental strategy, including the study of germ-free versus specific pathogen-free mice, colonization conditions, probiotic bacteria administration, faecal microbial transplant or antibiotic treatment. Up or down arrows indicate a respective increase or decrease in listed disease or function following microbial manipulation in the germ-free, probiotic or antibiotic state. Note that subtle variations in experimental methodology in faecal transplant studies mean that the effects shown here are likely to be an oversimplification (for example, transfer could be made from a donor mouse or human, from a control or symptomatic animal or individual and/or into a wild-type or a disease model recipient).

gut microbiota is sufficient to exacerbate neurological and psychological symptoms in some mouse models of multiple sclerosis³⁹, Parkinson disease⁴⁰, Alzheimer disease⁴¹, depression⁴², schizophrenia³³, attention-deficit hyperactivity disorder⁴³ and, possibly, autism spectrum disorder (ASD)⁴⁴. These studies are accomplished through the use of faecal microbial transplantation, whereby the faecal microbiota from human donors or other mouse models is used to colonize germ-free mice, limiting the confounding variables typical to human studies by using well-controlled, albeit reductionist, preclinical systems⁴⁵. Although these provocative studies are exciting, conclusions drawn from a small number of human donors remain speculative unless they are reproduced in larger cohorts of patients. In addition, causal inferences from human-murine microbiota transfer studies

are limited by inter-species differences in both microbiology and neurobiology⁴⁵. More established bacterial manipulations, such as treatment with particular bacteria or depletion of bacteria with antibiotics, have been shown to ameliorate disease symptoms in mouse models of ASD^{46,47}, multiple sclerosis^{48–50}, anxiety and depression^{15,51-53}, cognitive defects^{54,55} and Parkinson disease⁴⁰, as well as in humans with ASD^{56,57}, multiple sclerosis⁵⁸, anxiety and depression⁵⁹⁻⁶². Some of the effects of bacterial treatments on human brain activity have been characterized by changes observed in functional magnetic resonance imaging^{63,64}. Thus, emerging evidence suggests that neurological states may be impacted by gut microorganisms and their by-products.

Various associations between altered microbiome profiles and diseases of

the brain have been described, and the contribution of microbial communities or particular bacterial species to behaviour, cognition and neurodegeneration is continually being established⁶⁵. Furthermore, the gut microbiome harbours astonishing genetic diversity, with more than 22 million genes sequenced from human gut microbial populations, and an immense pool of unique enzymes capable of producing and modifying a wide array of chemical structural groups⁶⁶. We build on these foundational discoveries to describe and conceptualize how decoding of chemical messages that mediate the observed effects of the gut-brain axis provides promise in both understanding and treating a number of neurologic diseases. The following sections will delineate categories by source of the precursor (de novo bacterial, host or diet-derived sources) that can be transformed by gut bacteria and the bioactive molecules that result from microbial metabolism (FIG. 2). Brief descriptions of the effects of specific molecules are also provided.

Production of bacterial molecules

Microorganisms produce many proteins, vitamins and structural components that can serve to benefit or negatively affect the host. Many of these are generated via multistep biosynthetic pathways otherwise absent in mammals⁶⁷. These molecules sustain bacterial functions, such as signalling, structural components and energy sources, although some, such as proteinaceous toxins, are mainly known for their roles affecting host systems.

Microorganism-associated

molecular patterns. Microorganismassociated molecular patterns (MAMPs) are well-conserved components of microbial cells, and they are acutely detected by the host throughout the body, including the brain68. MAMPs play crucial roles in structural integrity and basic function for all classes of microorganisms and are complex molecules comprising diverse chemical groups including nucleotides, lipids, carbohydrates and peptides⁶⁹. The absence of MAMPs in germ-free mice leads to incomplete immune and neurodevelopment, but their presence can induce acute or chronic inflammation associated with various neurological disorders if the host response to MAMPs remains elevated or unchecked70. Two principle cellular surface component MAMPs that appear to be sufficient to

alter brain development and function are peptidoglycan and lipopolysaccharide (LPS). Peptidoglycan, a structural component of almost all bacterial cell walls, was recently shown to be translocated into the developing brain, affecting gene expression and social behaviour⁷¹. LPS, another ubiquitous surface molecule of Gram-negative bacteria, has been detected co-localizing with its receptor in rat brains72. LPS injection induces sickness behaviour⁷³, cognitive impairment⁷⁴ and acute depressive-like behaviours in mice75 and affects fetal brain development^{76,77}. Additionally, chronic or acute exposure to MAMPs is used to promote disease-related symptoms in models of ASD, depression, Parkinson disease and synucleinopathy78-80. These conserved microbial molecules may regulate mammalian behaviour through immune-mediated pathways via direct sensing by receptors expressed in the brain or activation of systemic inflammation and cytokine production, which can lead to altered neurological function and neuronal stress or cell death⁸¹. The presence, structure

and immunomodulatory activity of MAMPs vary between species of bacteria, and shifts in the gut community could, therefore, affect the level of exposure and response of the host to particular MAMPs, which in turn can influence downstream health status and behaviour.

Toxins. Proteinaceous toxins produced by some bacteria exert negative effects on the host nervous system. These toxins are often similar in general structure, with multiple subunits that activate cell-surface or intracellular receptors, and can be produced by opportunistic pathogens that may reside in the commensal community for long periods of time without causing overt disease in the gut or the brain⁸². Several species of Clostridia are known to produce many toxins, such as lethal toxin, toxin B, epsilon toxin and enterotoxin, that can reach the brain through the systemic circulation, disrupt and cross the blood-brain barrier (BBB), inhibit neurotransmitter release and/or decrease

neuron viability in targets ranging from the gut to the hippocampus^{82–86}. *Staphylococcus* spp. and *Bacillus* spp. produce toxins, staphylococcal enterotoxins and cereulide, that stimulate the vagus nerve, sending signals to the brain and inducing vomiting and sickness behaviour^{87–89}. Other species, such as *Salmonella* spp. and *Escherichia* spp., produce a class of proteins called amyloids, which aggregate in the intestine and may spread to the brain with a prion-like disease pattern and may contribute to neurodegeneration, such as in Parkinson disease and Alzheimer disease^{90–93}.

Transformation of host metabolites

Constant metabolic flux is sustained across the intestinal epithelial barrier as nutrients are absorbed and waste is secreted. The microbiota is exposed to and chemically interacts with many host molecules. The two classes of host-derived metabolites with the most evidence for gut-brain interactions are bile acids and steroid hormones.

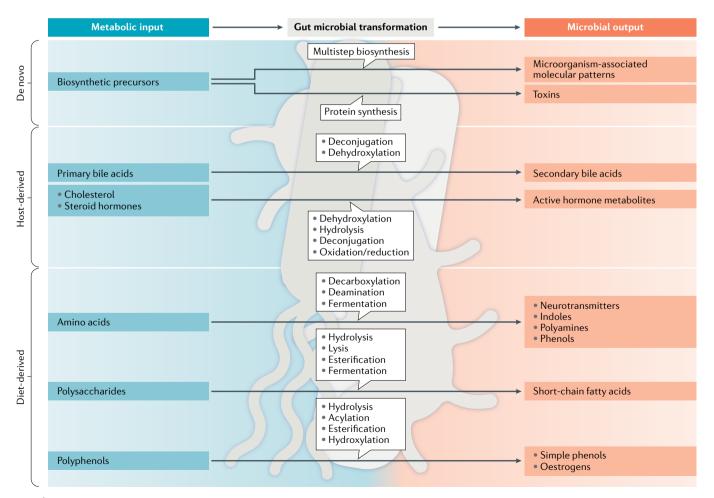


Fig. 2 | Gut bacterial metabolites. The modification of various substrates by the gut microbiota is shown broken down into three categories (left) of de novo bacterial, host-derived or dietary molecules. These substrates (metabolic input, shown in blue) are metabolized by many chemical processes by the microbiota (general examples are shown in white boxes). Many of the resulting metabolites have been shown to affect the brain (shown in red).

Bile acids. Primary bile acids are products of host cholesterol metabolism that play a major role in fat digestion and signalling in energy metabolism, even in the brain⁹⁴⁻⁹⁷. Circulating bile acids can cross the BBB and may act directly on their receptors in the brain, or have a more indirect effect by activating gut receptors, leading to the release of signals such as fibroblast growth factor and glucagon-like peptide 1, which can influence neuronal activity in multiple brain regions or the vagus nerve⁹⁸. The most common primary bile acids are cholic acid and chenodeoxycholic acid, and these are often conjugated with the amino acid glycine or taurine⁹⁹. Many gut bacterial species help maintain cholesterol homeostasis by modifying primary bile acids into secondary bile acids through dehydroxylation by dehydratase enzymes, deconjugation of the amino acid groups with bile salt hydrolases and further degradation with other enzymatic machinery^{99,100}. Bacterial modification changes the signalling of bile acids on membrane and nuclear receptors, and alters their solubility and circulation⁹⁴. Regulation of the presence and clearance of bile acids is involved in proper brain function, as defects in these pathways lead to many neurological phenotypes in mice and humans, such as demyelination, motor dysfunction, neuroinflammation, seizures and learning impairment^{96,101-112}. Bacterial influences on bile acid conjugation and levels may be influencing these brain phenotypes. For instance, alterations in bacterial-associated bile acid levels have been observed in human and mouse model studies of Parkinson disease¹¹³, Alzheimer disease114,115, multiple sclerosis116,117, alcohol dependency¹¹⁸ and ASD¹¹⁹, and bile acids are known to affect the hypothalamicpituitary-adrenal axis^{111,112,120}. In fact, all secondary bile acids produced by bacteria are detected in the brain of patients with Alzheimer disease, and increased ratios of secondary bile acids are correlated with their cognitive impairment and changes in brain imaging^{114,115}. Some bile acids are even used as potential treatments for neurological issues such as amyotrophic lateral sclerosis and stroke^{108,121,122}. The presence of bacteria in the gut changes host-wide bile acid levels, and community changes in the gut microbiota influence the levels and properties of bile acids¹²³⁻¹²⁹. These changes could be advantageous or detrimental. The most mechanistic link known between microbial metabolism of bile acids and potential neurological function may be that levels of deoxycholic acid directly increased by the microbiota are sufficient to induce

production of the major neurotransmitter, serotonin, in gut enterochromaffin cells¹³⁰. Gut serotonin levels may affect brain function in ways that are yet unknown, as hippocampal levels of serotonin are affected by the microbiota in mice, but any possible further links between gut and brain serotonin levels are not clear⁸. Cause and effect relationships between microbial manipulation of bile acids and brain function remain to be clearly defined.

Steroid hormones. Steroid hormone signalling is crucial for proper brain structure development, cognition, memory, decision-making and sexual behaviours, as well as playing a role in protection from social isolation and depression-like phenotypes¹³¹⁻¹³⁷. Up to 15% of some of these hormones produced daily are detectable in the gut, as they circulate through the body, bringing them into contact with the microbiota^{138,139}. The gut microbiota can influence levels of some hormones by shifting the ratio of active and inactive steroid levels through different degradation and activation pathways¹⁴⁰⁻¹⁴². The two best-studied classes for which this is the case are androgens and oestrogens. In many cases, hormones can be conjugated for excretion, and bacteria can remove the conjugation group with hydrolytic enzymes such as β-glucuronidases (GUSs) and β -glucosidases, which reactivate the molecule for continued circulation and activity¹⁴³⁻¹⁴⁶. Members of the microbiota can also convert cholesterol to androgens147,148, activate pro-androgens149,150 and metabolize testosterone into other potent androgens^{151,152}. Oestrogens are broken down in oxidative and reductive reactions in human faecal samples144,153-155. In fact, the term 'estrobolome' has been coined to describe the large collection of enteric bacterial genes capable of metabolizing oestrogens¹⁵⁶. Shifts in the gut microbiota and steroid hormone levels are associated with each other in postmenopausal women^{157,158} but, although the potential capacity for microbial metabolism of host hormones is vast, direct effects on brain function remain largely untested. If microbially influenced oestrogens do have direct neurological effects, they are likely neuroprotective, as oestrogens have antiinflammatory effects on microglia¹⁵⁹, and lowered levels of oestrogens due to altered microbial communities appear to increase cognitive impairment and chronic inflammation^{160,161}. Microorganisms may be sufficient for these phenotypes, as some steroid hormone levels can be transferred by

microbial faecal transplant between mice¹⁶², but further work is required to directly link hormone metabolites produced by the gut microbiota to neurological disease.

Transformation of dietary metabolites

The composition of the gut microbiota is heavily dependent on the dietary input of the host^{3,163}. The frequency of meals and types of foods influence the quantity of substrates metabolized by bacteria, which bacterial species wax and wane in abundance and, ultimately, the type and amount of downstream bacterial metabolites that are produced. Further, significant evidence shows that microbial metabolites of amino acids, complex plant polysaccharides and polyphenols exert an influence on the brain.

Amino acids. Microorganisms encode genetic machinery to produce many amino acids, some of which can contribute to circulating host levels^{164,165}. However, it is more likely that any microbial influence on the CNS via amino acid levels occurs through modification of dietary amino acids by deamination and decarboxylation pathways. The by-products of bacterial amino acid metabolism include ammonia, short-chain fatty acids (SCFAs), simple phenols, indole derivatives, neurotransmitters, organic acids, gaseous compounds and amines. Those most likely to affect brain function are described below.

Gut bacteria encode multiple gene pathways that metabolize the aromatic amino acids tyrosine, phenylalanine and tryptophan into a large group of downstream products, many of which are neurotransmitters^{166,167}. Tyrosine is metabolized to tyramine and then into two catecholamines, dopamine and noradrenaline. Tyramine in the intestine of germ-free mice also induces the production of serotonin¹³⁰. Noradrenaline is produced by gut bacteria^{12,168}, but it is not well understood how¹⁶⁸. However, multiple bacteria have been shown to synthesize noradrenaline up to the millimolar range in vitro^{169,170}. Catecholamine production by the microbiota may be sufficient for behavioural alterations, as mice treated with antibiotics were more sensitive to the dopamine signalling and behavioural effects of cocaine¹⁷¹. Whether these neuroactive molecules influence the local enteric nervous system or can affect the brain, even indirectly, is an active area of study.

Tryptophan is broken down by the microbiota into indole derivatives and also tryptamine and kynurenine metabolites, all of which have neuroactive

properties¹⁷²⁻¹⁷⁴. Some of these seem to only be produced by the microbiota, as they are undetectable in germ-free mice until bacterial colonization^{166,167}. Many of these can cross the BBB, and thus circulating tryptophan metabolites originating in the gut can contribute to levels in the brain¹⁷⁵. Indole derivatives such as indolepropionic acid have antioxidant properties, making this an attractive target for Alzheimer disease, whereas others such as indoxyl sulfate induce neuroinflammation in models of chronic kidney disease^{176,177}. Kynurenine metabolites act on neuronal glutamate receptors and affect memory, anxiety-like and stress-like behaviours¹⁷⁵. In fact, germ-free versus specific pathogen-free mice respond differentially in behavioural tests used to model depression-like phenotypes after depletion of dietary tryptophan (and, thus, all tryptophan microbial metabolites)27.

Besides neurotransmitters, tyrosine can also be metabolized by the microbiota into other simple phenols such as 4-ethylphenol or *p*-cresol. These metabolites are rapidly sulfated by the host to 4-ethylphenyl sulfate (4EPS) or *p*-cresyl sulfate, respectively. 4EPS is elevated in a mouse model of ASD and schizophrenia, as well as in samples from children with ASD¹⁷⁸, and was shown to be sufficient to cause an anxiety-like phenotype when injected into wild-type mice⁴⁶. *p*-Cresyl sulfate has been identified as a potential urinary biomarker for young children with ASD, and is correlated with altered oligodendrocyte markers in a mouse model of social and depressive-like behaviours, although these findings currently remain correlative^{179,180}.

Another amino acid affected by gut microorganisms is the major excitatory neurotransmitter glutamate, which is metabolized by a bacterial glutamate decarboxylase system to become the major inhibitory neurotransmitter GABA13,181,182. GABA can be further metabolized by bacteria to succinate by GABA aminotransferase and succinic semialdehyde dehydrogenase. Furthermore, metabolites either produced or influenced by the microbiota that affect the host GABA system have also been identified, such as γ -glutamyl amino acids, whose lowered levels are the mediators of diet-induced improvements in a mouse model of seizures¹⁸³. GABAproducing bacteria have been shown to alleviate depression-like and anxiety-like behaviours in mouse models⁵¹, and a strain engineered to produce GABA was sufficient to reduce sensitivity to visceral pain in rats¹⁸⁴. A GABA-producing microbiota is negatively

associated with depression in patients¹⁸⁵, and abnormalities in the glutamate/GABA circuits in the brain have been hypothesized as key in anxiety disorders, major depressive disorder, bipolar disorder, schizophrenia and ASD^{186–189}.

Arginine can be metabolized to four polyamines by the microbiota, which are present in all mammalian cells and play roles in many general processes of cell growth and differentiation, as well as regulating synaptic plasticity and memory formation via glutamate receptors¹⁹⁰. These polyamines are generated sequentially from arginine to agmatine, then putrescine, followed by spermidine and, then, spermine^{191,192}. Agmatine is a ligand for α_2 -adrenergic and imidazoline receptors in the brain¹⁹³⁻¹⁹⁶. Dysregulation of the polyamine system has been implicated in mood disorders, depression and Alzheimer disease, and polyamines have been studied as preclinical therapeutics for depression and anxiety-like behaviours, cognitive decline and drug dependency¹⁹⁷⁻²⁰³. As most mammalian neurotransmitters are derived from amino acid precursors, we speculate that bacterial transformation of amino acids into molecules that affect behaviour may represent a renewed microbial endocrinology focus in neuroscience²⁰⁴ that is worthy of further study.

Complex plant polysaccharides. Dietary fibre, made of complex carbohydrate polysaccharides, is not digested by the

Glossary

Bile acids

Complex lipid products of host cholesterol metabolism that play a major role in fat digestion and signalling in energy metabolism. Host bile acids (primary bile acids) are commonly modified by bacteria into secondary bile acids.

Enterochromaffin cells

Neuroendocrine cells in the gut lining that aid in gastrointestinal motility and produce 90% of the body's serotonin in response to persistent intestinal signals.

Germ-free mice

Mice reared in conditions completely absent of microbial exposure.

Gut microbiota

An intestinal community comprising bacteria and other microorganisms including viruses, fungi, protists and archaea that permanently or transiently inhabit the lower gastrointestinal tract, especially the small intestine and colon.

Microorganism-associated molecular patterns

(MAMPs). Well-conserved components of microbial cells that are acutely detected by the innate immune system of the host throughout the body, including the brain.

host and reaches the colon, where it is fermented by intestinal microorganisms with a diverse class of glycoside hydrolases and polysaccharide lyases into millimolar levels of SCFAs^{205,206}. SCFAs, mainly butyrate, propionate and acetate, are a rich energy source for colonic epithelial cells, and the remaining pool enters the systemic circulation where they may subsequently influence neurological function and development, seemingly for better or for worse, depending on the context. For instance, SCFAs were sufficient to exacerbate motor symptoms in a germ-free Parkinson disease mouse model⁴⁰, but they improved recovery from an experimental stroke mouse model²⁰⁷. Acetate has been shown to cross the BBB in mice and reduce feeding behaviours^{208,209}. Propionate protects the BBB through signalling via the G proteincoupled receptor (GPCR) FFAR3 (REF.²¹⁰) and improves multiple sclerosis symptoms in patients²¹¹, but injections of propionate have also been used to induce a rodent model of ASD^{212,213}. Butyrate is a potent inhibitor of histone deacetylases (HDACs), which regulate epigenetic signals of gene activation. As lowered histone acetylation is a characteristic of multiple neurodegenerative diseases²¹⁴, the pharmacological use of butyrate has been widely explored. Some preliminary success for butyrate treatment has been seen in beneficially lowered inflammation in mouse models of Huntington disease, Parkinson disease, ischaemic

Polyphenols

A vast class of thousands of plant-derived molecules containing at least one phenol group that are generally poorly absorbed by the host until being transformed by the gut microbiota into bioavailable and bioactive metabolites.

Short-chain fatty acids

(SCFAs). Fatty acids with chains of fewer than six carbons that are the end product of bacterial fermentation of complex polysaccharides and serve as energy source and signalling molecule in the host.

Specific pathogen-free mice

Mice conventionally colonized with a complete gut microbiota.

Steroid hormones

Circulating signalling molecules derived from cholesterol with an organic chemical structure consisting of four carbon rings and various regulatory roles in the host.

Vagus nerve

A principle neuronal connection between the gut and brain, comprising a bundle of neurons that sends and receives signals directly between gut tissue (and other organs) and the brainstem. These signals can then be further transmitted throughout the brain.

stroke, Alzheimer disease and memory impairment^{215–224}.

Polyphenols. Polyphenols comprise a vast class of thousands of plant-derived molecules containing at least one phenol group, and are being extensively studied as therapeutics for neurological disease²²⁵. Most polyphenols are generally poorly absorbed until being transformed by the gut microbiota into bioavailable and bioactive metabolites^{226,227}. Bacterial hydrolysis, acylation and/or esterification is followed by host modification by methylation, sulfation, hydroxylation or glucuronidation before these metabolites re-enter the gastrointestinal tract or reach other peripheral tissues^{228,229}. Phenolic metabolite levels are known to be altered in the brain after oral administration of parent polyphenols^{230–232}. Specific bacterial metabolites of oral polyphenol treatment that were measured in the brain, such as 3-hydroxybenzoic acid and 3-(3'-hydroxyphenyl)propionic acid, were shown to be capable of inhibiting hallmark amyloid aggregation and slowing progression of Alzheimer disease pathophysiology^{233–235}. Polyphenols were also protective against stress-induced depression-like behaviours via decreased inflammation and modulated synaptic plasticity through metabolites such as quercetin-3-O-glucuronide and malvidin-3-O-glucoside236. One polyphenol, ferulic acid, is liberated into circulation by gut microorganisms harbouring the ferulic acid esterase gene²³⁷. Ferulic acid administration stimulates neurogenesis in a corticosterone-treated depression mouse model, and is protective in mouse models of Alzheimer disease and cerebral ischaemia²³⁸⁻²⁴⁰. Polyphenols in treatments such as grape seed extract and resveratrol show promise in treating neuropathology and cognitive defects in mouse models of Alzheimer disease, Parkinson disease and tauophathies^{233,235,241-244}, but further tests with pure polyphenols are needed. Recently, it was shown that plant-derived epigallocatechin gallate can prevent motor symptoms induced by specific gut bacteria in a model of Parkinson disease²⁴⁵. Some polyphenols are phytoestrogens, which are metabolized by gut bacteria into equol and enterolactone derivatives²⁴⁶⁻²⁴⁹. Phytoestrogen metabolites can be either agonistic or antagonistic to oestrogen receptors and may have an impact on the neuroprotective pathways activated by classic oestrogen receptor ligands, although this structural class is large and heterogeneous, and direct effects on the brain remain to be conclusively shown^{250,251}.

Fig. 3 | Mechanistic examples of the routes of gut-brain communication. There are several different ways in which gut microbial metabolites can influence brain function. Specific examples are shown of vagus nerve modulation, enterochromaffin cell modulation, direct brain exposure and immunemediated communication. In the vagus nerve modulation panel, an example is shown where dietary tryptophan is converted to indole when mice are monocolonized by an Escherichia coli strain expressing the TnaA tryptophanase enzyme (TnaA + E. coli) compared with monocolonization with the control E. coli strain lacking TnaA, where no indole is produced. Bacterial modification of tryptophan was shown to result in activated vagal neurons and increased anxiety-like and depression-like behavioural phenotypes in the animals. In the enterochromaffin cell modulation panel, an example is shown where spore-forming bacteria from the Clostridiaceae and Turicibacteraceae families produce various metabolites including secondary bile acids, amino acid metabolites and short-chain fatty acids (SCFAs), which induce serotonin production by enterochromaffin cells and lead to elevated levels of circulating serotonin. We speculate that these serotonin levels may influence vagal nerve activity and/or brain levels of serotonin, denoted with a question mark in the figure. In the third panel, direct brain exposure via circulation, an example is shown where parent polyphenols, such as those found in grape seed extract, are modified by the microbiota into various phenolic metabolites that can be subsequently measured in the brain in association with reduced amyloid plaques and improved cognition. In the final panel (bottom right), immune system mediated, examples are shown where, on the left, a healthy diet including complex polysaccharides is fermented into SCFAs by the microbiota, which play important roles in G protein-coupled receptor (GPCR) signalling, histone deacetylase (HDAC) inhibition and lowered systemic inflammation that lead to decreased neuroinflammation. On the right, unbalanced microbiota can lead to altered levels of inflammatory bacterial lipopolysaccharide (LPS), which can lead to elevated neuroinflammation and depression-like behaviour via directly entering the brain or by inducing an elevated systemic immune response.

Other metabolites. Microbial GUS enzymes in the intestines remove glucuronide groups that mark metabolites for excretion by the host. As a result, the microbiota restores the original molecule and facilitates reuptake of the molecule back into the bloodstream^{252,253}. This process has been shown to directly regulate levels of many of the exogenous and endogenous compounds described herein^{168,254-256}.

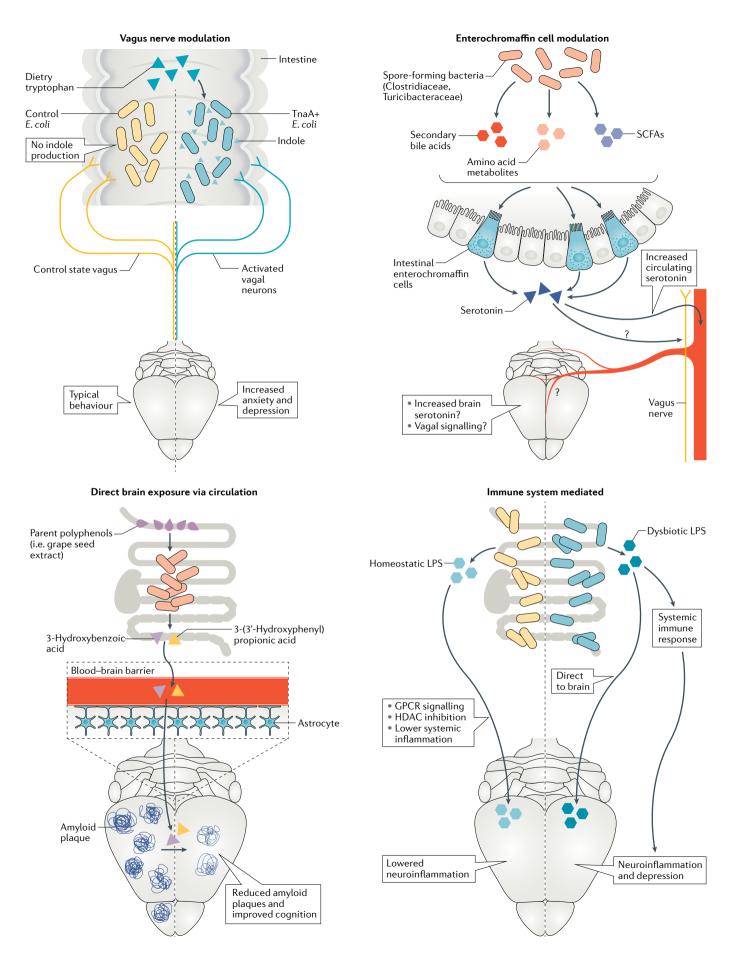
The gut microbiota also generates vitamins B and K^{67,257-261}, as well as unique lipid metabolites such as conjugated linoleic acids, hydroxy fatty acids and sphingolipids, several of which show biological activity in host health and disease and are known to be produced by particular bacterial species^{123,125-128,262-265}. Owing to the need for vitamins B and K during brain development, the high lipid content of the brain and the importance of lipids in signalling pathways, future work may illuminate connections of microbial lipid and vitamin metabolites with brain function.

Research into the production and function of bacterial metabolites has established that active chemical messaging occurs from the gut to the brain. Other bacterial molecules could have as yet undefined neuroactive properties, including any of the thousands of recently identified (but still uncharacterized) short peptides from the gut microbiota²⁶⁶. Given that identifying and characterizing the small molecules and peptide repertoires produced by the microbiota is a relatively new endeavour, it is likely that further neuroactive microbial metabolites will continue to be discovered. Defining mechanisms of action may lead to various health applications.

Routes of microbiota-brain signalling

Conduits of communication from the gut microbiota to the brain include activation of the vagus nerve, stimulation of endocrine cells (including enterochromaffin cells), immune-mediated signalling and transport of gut-derived metabolites from the circulation into the brain. All routes comprising the gut-brain axis are thought to be co-opted by the microbiota to impact brain activity and behaviour, and signalling through any one of them may be intertwined with other routes (FIG. 3).

Vagus nerve activation. The vagus nerve directly links the muscular and mucosal layers along the gastrointestinal tract to the brainstem and is a well-established signalling pathway affecting feeding, anxiety-like, depressive-like and social behaviours^{51,267,268}. Enteric pathogens and probiotics affect these behaviours through activation of vagal neurons, which then alters downstream neurological activity, including altered BDNF, GABA and oxytocin signalling in the brain^{51,267–269}. These responses are ablated following vagotomy, which severs the vagus nerve, but the specific bacterial metabolites mediating these effects remain largely unidentified. One recent study did measure the effects of a specific metabolite through vagus signalling, although additional routes of signalling could also be involved¹⁷³. In this work, rats were mono-colonized with either an Escherichia coli strain that converts



dietary tryptophan into indole with the *tnaA* tryptophanase or a mutant *E. coli* deficient in indole production. Rats exposed to indole in the gut displayed increased anxiety-like and depressive-like behaviours and activated vagal neurons¹⁷³.

Enterochromaffin cell stimulation.

Enterochromaffin cells are endocrine cells in the gut lining that produce and secrete 90% of the body's serotonin in response to persistent intestinal signals²⁷⁰. Enterochromaffin cell production of serotonin impacts its circulating levels^{130,167,271,272} and has the potential to influence brain activity directly or indirectly. Improved performance in mouse models of depression have been shown by probiotic treatment with *Bifidobacterium* spp. in a study that concurrently observed an increase in serotonin levels in the brain and also increased secretion of the serotonin precursor in enterochromaffin cells in vitro. However, no mechanistic connection between bacterial treatment, potential serotonin regulation and depressive-like phenotypes has been proved²⁷³. Colonic enterochromaffin cells do express receptors for, and respond to, various microbial metabolites, including MAMPs, SCFAs, aromatic amino acid metabolites and secondary bile acids²⁷⁴⁻²⁷⁸. One bacterial subset recently identified to greatly promote serotonin biosynthesis from enterochromaffin cells are sporeforming bacteria such as Clostridia spp. A collection of metabolites made by these bacteria in vivo were shown to be sufficient for serotonin-induction activity in vitro, including a-tocopherol, butyrate, cholate, deoxycholate, *p*-aminobenzoate, propionate and tyramine¹³⁰. A subset was individually tested with temporal intestinal administration as well, and deoxycholate, a-tocopherol, *p*-aminobenzoate and tyramine were each sufficient to induce production of serotonin by enterochromaffin cells¹³⁰. Interestingly, recent work showed that oral administration of a selective serotonin reuptake inhibitor, which increases bioavailability of gut serotonin and is used to treat depression, may be dependent on vagus nerve activation for its improvement of depressive-like behaviour in mice²⁷⁹. This supports the possibility that enterochromaffin cell production of serotonin has the potential to relay signals beyond the gut and reach the brain, possibly by intersecting with other known routes of gut-brain signalling in both developmental and acute contexts.

Immune-mediated signalling. The gut microbiota provides cues for the maturation of the neuroimmune system, and a loss of these cues during development results in lifelong dysfunction of this system²⁸⁰. However, chronic exposure to inflammation driven by shifts in gut microbiota and increased intestinal permeability may also factor into various neurological diseases70. Bacterial metabolites that serve as MAMPs, such as LPS, have been used to activate the immune system in models of ASD and schizophrenia, and also induce depression-like symptoms in mice^{75,76,281}. Other gut metabolites likely dampen chronic inflammation. SCFAs, for example, interact closely with the immune system through activation of GPCRs and inhibition of HDAC activity. A high-fibre diet, leading to higher levels of SCFAs, results in lower levels of circulating proinflammatory cytokines^{282,283}. Activation of GPCRs (FFA2 and GPR109a) by SCFAs can inhibit inflammatory signalling pathways, and HDAC inhibition by SCFAs, especially butyrate, leads to lowered inflammation in vivo^{117,284-286}. These examples likely represent initial discoveries into the potential impacts of microbial molecules on neuroimmune signalling.

Direct transfer of metabolites to the brain. Many microbial metabolites produced in the gut can pass into systemic circulation at varying levels and rates. One example is the polyphenolic metabolite group, where recent studies have shown that parent polyphenols are virtually undetectable in the bloodstream or urine, but that bacterial metabolites produced from polyphenol precursors enter circulation at levels sufficient to exert biological effects^{287,288}. In fact, the brain appears to be a major target for some polyphenolic microbial metabolites^{289,290}. Although in vivo evidence remains lacking, in vitro cultures have shown that polyphenol metabolites are able to cross BBB model systems and exert protective effects on neuronal cultures, mostly through a decrease in inflammatory responses^{291,292}. Furthermore, derivatives of oral polyphenolic treatment were measured in the blood and brain of rats and were found to decrease aggregation of neurotoxic aggregates and promote neuroplasticity^{231,233,293}.

Although gut-brain connections are well established, clear mechanistic details of the bacterial molecules working through each conduit are still limited. Understanding how the microbiome signals from the gut to the brain may provide insights into rational drug discovery platforms directed to targets in the gastrointestinal tract, which may overcome current challenges in the delivery of drugs to targets in the brain.

Cell-specific effects in the brain

Studies continue to build on the foundational understanding of the gut–brain axis to explore which cells in the brain are affected directly or indirectly by specific bacterial metabolites. Much work is needed to systematically demonstrate that these chemical messengers derived from gut bacteria influence the development or function of specific brain cells. Here, we summarize the current evidence that gut microbial metabolites may affect cells in the brain (FIG. 4).

Neurons. As the primary signalling cell of the brain controlling behaviour, neurons may, in essence, be the ultimate target affected by every metabolite described in this Perspective. All unidentified metabolites exerting the effects of bacterial communities that influence the vagus nerve probably activate neurons. More specifically, neurotoxins provide a stark example of bacterial molecules affecting neurons. Some neurotoxins are produced by commensal members of the microbiota and exert local or CNS effects to dysregulate or kill neurons^{82,83,87–89}. The microbiota also produces or induces the production of neurotransmitters and their precursors, including serotonin, adrenaline, GABA, histamine, acetylcholine, glutamate and dopamine, which could dramatically affect the balance of excitatory and inhibitory neurotransmission in enteric, vagal, peripheral and central neurons²⁹⁴. Neurons also express pattern recognition receptors, and activation of these receptors has been shown to regulate neuronal differentiation, proliferation and axon generation as well as neuroinflammation. Some of this is likely due to host ligands, but MAMPs such as peptidoglycan are also detected in the brain and could be activating receptors such as TLR2, PGLYRP2 or NOD1, which are expressed in neurons, through similar mechanisms^{71,295-297}. Neurons are also influenced by SCFAs, as acetate enters the brain and activates neurons in the hypothalamus^{208,209}. Finally, in vitro screens identified neuroactive molecules produced by gut microorganisms, such as quorum sensing molecules, that affect the viability, morphology, differentiation and inflammatory responses of neurons²⁹⁸. Although the latter need to be validated

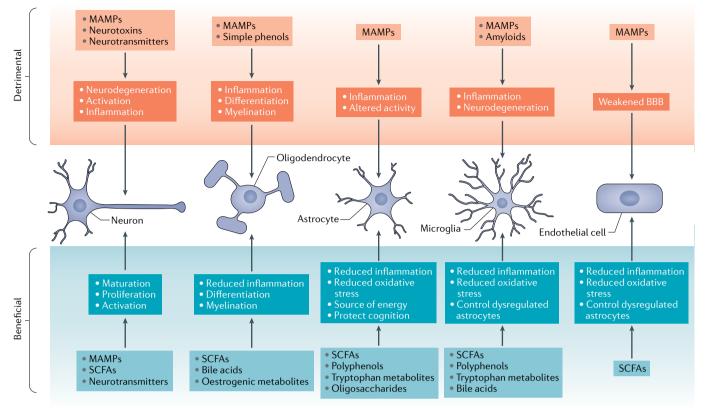


Fig. 4 | **Brain cell-specific effects of microbial metabolites.** Some microbial metabolites have known cellular targets in the brain. The beneficial and detrimental effects of these interactions are summarized. BBB, blood–brain barrier; MAMPs, microorganism-associated molecular patterns; SCFAs, short-chain fatty acids.

in vivo, they illustrate the possibility of a vast amount of interface between neurons and microbial metabolites.

Astrocytes. Astrocytes provide support to other cells and repair damage in the brain. Metabolites, including specific oligosaccharides and polyphenols, SCFAs and tryptophan metabolites, can affect astrocyte function. Tryptophan metabolites modulate the aryl hydrocarbon receptor in astrocytes and affect their activity by decreasing their inflammatory state and altering their interaction with microglia²⁹⁹⁻³⁰². Polyphenolic metabolites and pure SCFAs such as butyrate have in vitro effects on astrocytes, and have been shown to decrease neuroinflammation and oxidation³⁰³⁻³⁰⁵. The SCFA acetate is used as an energy source by these cells in the brain²⁰⁹. Oligosaccharides and polyphenols such as those from the plant Morinda officinalis, which are metabolized by bacteria into SCFAs and other lipid derivatives, have been shown to have protective effects in Alzheimer disease through astrocyte function³⁰⁶. Astrocytes also express G protein-coupled bile acid receptor 1 (TGR5), which can be activated by bile acids with a resulting decrease in

neuroinflammation, and may be relevant to hepatic encephalopathy⁹⁶.

Oligodendrocytes. Oligodendrocytes produce the myelin that insulates neuronal axons, with dynamic crosstalk between the two cell types even throughout adulthood. Metabolite effects on oligodendrocyte proliferation, differentiation and function could have widespread effects on neurological health. In the mouse model of the demyelinating disease multiple sclerosis, therapeutic gut microbiota manipulations have been successful and are accompanied by changes in metabolomic profiles associated with alleviated disease symptoms^{50,307–312}. There is some evidence that improvements may be due to a decrease in inflammatory LPS levels, an increase in SCFAs and an altered profile of bile acids, although whether direct activity on oligodendrocytes occurs or whether they indirectly benefit from lowered inflammation has not been elucidated^{129,308}. In vitro, the bacterial phenolic metabolite p-cresol may directly impair oligodendrocyte maturation and myelin production¹⁸⁰. Another class of molecules known to affect oligodendrocyte differentiation and myelination are oestrogenic

molecules³¹³⁻³¹⁷. Microorganisms do modify many oestrogenic metabolites¹⁵⁶, but a conclusive link between in vivo microbial production of these metabolites and oligodendrocytes has not yet been proved.

Endothelial cells. Blood vessels are lined with endothelial cells, which are the major cell type responsible for maintenance of the BBB that largely determines molecular entry into the brain³¹⁸. Modulation of BBB permeability by microbial metabolites could greatly alter uptake of drugs, host molecules and other gut metabolites, but concrete examples of this mechanism remain elusive. For example, bacterial metabolites such as LPS from some bacterial species increase permeability in vivo in a dose and bacterial strain-dependent manner³¹⁹, and germ-free mice appear to have a leakier BBB than conventional mice³²⁰. LPS stimulation of endothelial cells can also lead to cerebral cavernous malformations, which in turn lead to seizures and strokes³²¹. SCFAs have been shown to decrease permeability of the BBB through activating SCFA receptors expressed in endothelial cells and concurrent increases in expression of tight junction proteins that seal these cells into a successful barrier^{210,320}.

Microglia. The primary immune cells in the brain are known as microglia, and as such are responsible for much of the damage associated with neuroinflammation in diseases such as Parkinson disease and Alzheimer disease³²². It is not surprising, then, that pro-inflammatory signals from MAMPs induce mature and cytokineproducing microglia whereas the generally anti-inflammatory cues from polyphenolic, SCFA and bile acid metabolites work via microglia to lower oxidative stress in the brain^{286,323-325}. However, the effects of some of these signals on microglia are complex, as SCFAs, and probably other microbial signals, exacerbate symptoms in a germfree mouse model of Parkinson disease⁴⁰. Another recent work discovered that microbial tryptophan metabolites such as indoxyl-3-sulfate control the activation of microglia, which in turn alter the behaviour of astrocytes³⁰⁰.

Although examples of cell-specific effects by the microbiome are both sparse and superficially described to date, these foundational studies represent critical steps in uncovering the underlying neuronal circuits, brain regions and systems-level connections of the gut microbiome-brain axis.

Perspectives

The gut microbiota, the gastrointestinal tract and the brain have historically been studied independently, but a growing appreciation for their interconnectedness may lead to transformative advances in biomedicine. Identifying and characterizing causal or contributing roles for particular microorganisms and microbial communities should be a primary focus of gut microbiome-brain research. However, the current state of the field remains largely correlative with descriptions of gut metabolite profiles in the context of neurologic states, whereas specific examples for effects by gut-derived molecules on brain cells, brain activity and behaviour are limited to a handful of studies. Additional progress is also needed to further understand the physical pathways employed by the microbiome in mediating communication between the gut and the brain. The various routes of direct and indirect chemical signalling are not mutually exclusive, and some metabolites potentially exert effects on multiple conduits to the brain.

As specific effects of microbial molecular messages and their gut-brain signalling routes continue to be uncovered, the potential increases for development of novel therapeutic principles and modalities. Continued, rigorous distinction between the correlative and causative links connecting gut metabolites to the brain may lead the way for new hypotheses for disease aetiology and treatment. For example, dietary interventions to shift the microbial community in favour of bacteria capable of producing beneficial chemical signals, or away from those generating pathogenic compounds, can be envisioned. Understanding of the microbial molecules crucial for health could allow the deployment of specific probiotics for specific maladies, based on empiric evidence that is lacking in current commercially available probiotic strains. Importantly, a deeper understanding of the mechanistic underpinnings for gut-brain connections in neurological diseases could lead to a world with targeted therapeutics directed at microbial effectors. Drugs could selectively inhibit production of harmful metabolites by targeting specific bacterial enzymes, which are evolutionarily divergent from human enzymes, thus increasing the available therapeutics while decreasing off-target effects. Further, microbial metabolites themselves may be therapeutically administered. The merger of microbiome and neuroscience research offers the possibility of understanding the basic 'wiring' and functions of the gut-brain axis, and also provides potential opportunities for near-term, actionable advances in human health.

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Published online 16 October 2020

- Cho, I. & Blaser, M. J. The human microbiome: at the interface of health and disease. *Nat. Rev. Genet.* 13, 260–270 (2012).
- Koppel, N., Rekďal, V. M. & Balskus, E. P. Chemical transformation of xenobiotics by the human gut microbiota. *Science* **356**, eaag2770 (2017).
- Sonnenburg, J. L. & Bäckhed, F. Diet–microbiota interactions as moderators of human metabolism. *Nature* 535, 56–64 (2016).
- Nyangahu, D. D. & Jaspan, H. B. Influence of maternal microbiota during pregnancy on infant immunity. *Clin. Exp. Immunol.* **198**, 47–56 (2019).
- Human Microbiome Project Consortium. Structure, function and diversity of the healthy human microbiome. *Nature* 486, 207–214 (2012).
- Kagnoff, M. F. Immunology of the intestinal tract. *Gastroenterology* 105, 1275–1280 (1993).
- Furness, J. B., Callaghan, B. P., Rivera, L. R. & Cho, H.-J. The enteric nervous system and gastrointestinal innervation: integrated local and

central control. Adv. Exp. Med. Biol. 817, 39–71 (2014).

- Clarke, G. et al. The microbiome-gut-brain axis during early life regulates the hippocampal serotonergic system in a sex-dependent manner. *Mol. Psychiatry* 18, 666–673 (2013).
- Neufeld, K. M., Kang, N., Bienenstock, J. & Foster, J. A. Reduced anxiety-like behavior and central neurochemical change in germ-free mice. *Neurogastroenterol. Motil.* 23, 255–e119 (2011).
- Fröhlich, E. E. et al. Cognitive impairment by antibiotic-induced gut dysbiosis: analysis of gut microbiota-brain communication. *Brain Behav. Immun.* 56, 140–155 (2016).
- Desbonnet, L. et al. Gut microbiota depletion from early adolescence in mice: implications for brain and behaviour. *Brain Behav. Immun.* 48, 165–173 (2015).
- Diaz Heijtz, R. et al. Normal gut microbiota modulates brain development and behavior. *Proc. Natl Acad. Sci. USA* 108, 3047–3052 (2011).
- Matsumoto, M. et al. Cerebral low-molecular metabolites influenced by intestinal microbiota: a pilot study. Front. Syst. Neurosci. 7, 9 (2013).
- Sudo, N. et al. Postnatal microbial colonization programs the hypothalamic–pituitary–adrenal system for stress response in mice. *J. Physiol.* 558, 263–275 (2004).
- Savignac, H. M. et al. Prebiotic feeding elevates central brain derived neurotrophic factor, *N*-methyl-Daspartate receptor subunits and D-serine. *Neurochem. Int.* 63, 756–764 (2013).
- Bora, S. A., Kennett, M. J., Smith, P. B., Patterson, A. D. & Cantorna, M. T. The gut microbiota regulates endocrine vitamin D metabolism through fibroblast growth factor 23. Front. Immunol. 9, 408 (2018).
- Monteggia, L. M. et al. Essential role of brain-derived neurotrophic factor in adult hippocampal function. *Proc. Natl Acad. Sci. USA* **101**, 10827–10832 (2004).
- Luczynski, P. et al. Adult microbiota-deficient mice have distinct dendritic morphological changes: differential effects in the amygdala and hippocampus. *Eur. J. Neurosci.* 44, 2654–2666 (2016).
- Lu, J. et al. Microbiota influence the development of the brain and behaviors in C57BL/6J mice. *PLoS ONE* 13, e0201829 (2018).
- Hoban, A. E. et al. Regulation of prefrontal cortex myelination by the microbiota. *Transl Psychiatry* 6, e774 (2016).
- Hoban, A. E. et al. The microbiome regulates amygdala-dependent fear recall. *Mol. Psychiatry* 23, 1134–1144 (2018).
- Desbonnet, L., Clarke, G., Shanahan, F., Dinan, T. G. & Cryan, J. F. Microbiota is essential for social development in the mouse. *Mol. Psychiatry* 19, 146–148 (2014).
- Buffington, S. A. et al. Microbial reconstitution reverses maternal diet-induced social and synaptic deficits in offspring. *Cell* 165, 1762–1775 (2016).
- Leclercq, S. et al. Low-dose penicillin in early life induces long-term changes in murine gut microbiota, brain cytokines and behavior. *Nat. Commun.* 8, 15062 (2017).
- Luo, Y. et al. Gut microbiota regulates mouse behaviors through glucocorticoid receptor pathway genes in the hippocampus. *Transl. Psychiatry* 8, 187 (2018).
- Huo, R. et al. Microbiota modulate anxiety-like behavior and endocrine abnormalities in hypothalamic–pituitary–adrenal axis. Front. Cell. Infect. Microbiol. 7, 489 (2017).
- Lukić, I., Getselter, D., Koren, O. & Elliott, E. Role of tryptophan in microbiota-induced depressive-like behavior: evidence from tryptophan depletion study. *Front. Behav. Neurosci.* 13, 123 (2019).
- Ceylani, T., Jakubowska-Doğru, E., Gurbanov, R., Teker, H. T. & Gozen, A. G. The effects of repeated antibiotic administration to juvenile BALB/c mice on the microbiota status and animal behavior at the adult age. *Heliyon* 4, e00644 (2018).
- Zhai, B., Shang, X., Fu, J., Li, F. & Zhang, T. Rapamycin relieves anxious emotion and synaptic plasticity deficits induced by hindlimb unloading in mice. *Neurosci. Lett.* 677, 44–48 (2018).
- Hoban, A. E. et al. Behavioural and neurochemical consequences of chronic gut microbiota depletion during adulthood in the rat. *Neuroscience* 339, 463–477 (2016).
- Wang, B., Yao, M., Lv, L., Ling, Z. & Li, L. The human microbiota in health and disease. *Engineering* 3, 71–82 (2017).

- Kang, D.-W. et al. Differences in fecal microbial metabolites and microbiota of children with autism spectrum disorders. *Anaerobe* 49, 121–131 (2018)
- Zheng, P. et al. The gut microbiome from patients with schizophrenia modulates the glutamate–glutamine– GABA cycle and schizophrenia-relevant behaviors in mice. *Sci. Adv.* 5, eaau8317 (2019).
 Jiang, H. et al. Altered fecal microbiota composition in
- Jiang, H. et al. Altered fecal microbiota composition in patients with major depressive disorder. *Brain Behav. Immun.* 48, 186–194 (2015).
- Prehn-Kristensen, A. et al. Reduced microbiome alpha diversity in young patients with ADHD. *PLoS ONE* 13, e0200728 (2018).
- Vogt, N. M. et al. Gut microbiome alterations in Alzheimer's disease. *Sci. Rep.* 7, 1–11 (2017).
 Schenerians, F. et al. Gut microbiota are related
- Scheperjans, F. et al. Gut microbiota are related to Parkinson's disease and clinical phenotype. *Mov. Disord.* **30**, 350–358 (2015).
- Jangi, S. et al. Alterations of the human gut microbiome in multiple sclerosis. *Nat. Commun.* 7, 12015 (2016).
- Berer, K. et al. Commensal microbiota and myelin autoantigen cooperate to trigger autoimmune demyelination. *Nature* **479**, 538–541 (2011).
- Sampson, T. R. et al. Gut microbiota regulate motor deficits and neuroinflammation in a model of Parkinson's disease. *Cell* 167, 1469–1480.e12 (2016).
- Fujii, Ý. et al. Fecal metabolite of a gnotobiotic mouse transplanted with gut microbiota from a patient with Alzheimer's disease. *Biosci. Biotechnol. Biochem.* https://doi.org/10.1080/09168451.2019.1644149 (2019).
- Żheng, P. et al. Gut microbiome remodeling induces depressive-like behaviors through a pathway mediated by the host's metabolism. *Mol. Psychiatry* 21, 786–796 (2016).
- Tengeler, A. C. et al. Gut microbiota from persons with attention-deficit/hyperactivity disorder affects the brain in mice. *Microbiome* 8, 44 (2020).
- Sharon, C. et al. Human gut microbiota from autism spectrum disorder promote behavioral symptoms in mice. *Cell* **177**, 1600–1618 (2019).
- Walter, J., Armet, A. M., Finlay, B. B. & Shanahan, F. Establishing or exaggerating causality for the gut microbiome: lessons from human microbiotaassociated rodents. *Cell* **180**, 221–232 (2020).
- Hsiao, E. Y. et al. Microbiota modulate behavioral and physiological abnormalities associated with neurodevelopmental disorders. *Cell* **155**, 1451–1463 (2013).
- Tabouy, L. et al. Dysbiosis of microbiome and probiotic treatment in a genetic model of autism spectrum disorders. *Brain Behav. Immun.* **73**, 310–319 (2018).
- Ochoa-Repăraz, J. et al. Role of gut commensal microflora in the development of experimental autoimmune encephalomyelitis. J. Immunol. 183, 6041–6050 (2009).
- Seifert, H. A. et al. Antibiotics protect against EAE by increasing regulatory and anti-inflammatory cells. *Metab. Brain Dis.* 33, 1599–1607 (2018).
- He, B. et al. *Lactobacillus reuteri* reduces the severity of experimental autoimmune encephalomyelitis in mice by modulating gut microbiota. *Front Immunol* 10, 384 (2019).
- Bravo, J. A. et al. Ingestion of *Lactobacillus* strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. *Proc. Natl Acad. Sci. USA* **108**, 16050–16055 (2011).
- Liu, W.-H. et al. Genome architecture of *Lactobacillus* plantarum PS128, a probiotic strain with potential immunomodulatory activity. *Cut Pathog.* 7, 22 (2015)
- immunomodulatory activity. *Gut Pathog.* 7, 22 (2015).
 53. Burokas, A. et al. Targeting the microbiota-gut-brain axis: prebiotics have anxiolytic and antidepressant-like effects and reverse the impact of chronic stress in mice. *Biol. Psychiatry* 82, 472–487 (2017).
 54. Lee, H.-J., Lee, K.-E., Kim, J.-K. & Kim, D.-H.
- Lee, H.-J., Lee, K.-E., Kim, J.-K. & Kim, D.-H. Suppression of gut dysbiosis by *Bifidobacterium longum* alleviates cognitive decline in 5XFAD transgenic and aged mice. *Sci. Rep.* 9, 11814 (2019).
- Li, Y. et al. Neuroprotective effects of intravenous transplantation of bone marrow mononuclear cells from 5-fluorourcail pre-treated rats on ischemic stroke. *Behav. Brain Res.* 301, 287–292 (2016).
- 56. Sandler, R. H. et al. Short-term benefit from oral vancomycin treatment of regressive-onset autism. *J. Child. Neurol.* **15**, 429–435 (2000).
- 57. Kang, D.-W. et al. Microbiota transfer therapy alters gut ecosystem and improves gastrointestinal and

autism symptoms: an open-label study. *Microbiome* **5**, 10 (2017).

- Metz, L. M. et al. Trial of minocycline in a clinically isolated syndrome of multiple sclerosis. *N. Engl. J. Med.* **376**, 2122–2133 (2017).
- Allen, A. P. et al. *Bifidobacterium longum* 1714 as a translational psychobiotic: modulation of stress, electrophysiology and neurocognition in healthy volunteers. *Transl. Psychiatry* 6, e939 (2016).
- Tillisch, K. et al. Consumption of fermented milk product with probiotic modulates brain activity. *Gastroenterology* https://doi.org/10.1053/ j.gastro.2013.02.043 (2013).
- Steenbergen, L., Sellaro, R., van Hemert, S., Bosch, J. A. & Colzato, L. S. A randomized controlled trial to test the effect of multispecies probiotics on cognitive reactivity to sad mood. *Brain Behav. Immun.* 48, 258–264 (2015).
- Messaoudi, M. et al. Assessment of psychotropic-like properties of a probiotic formulation (*Lactobacillus helveticus* R0052 and *Bifidobacterium longum* R0175) in rats and human subjects. *Br. J. Nutr.* **105**, 755–764 (2011).
- Bagga, D. et al. Probiotics drive gut microbiome triggering emotional brain signatures. *Cut Microbes* 9, 486–496 (2018).
- Bagga, D. et al. Influence of 4-week multi-strain probiotic administration on resting-state functional connectivity in healthy volunteers. *Eur. J. Nutr.* 58, 1821–1827 (2019).
- Vuong, H. E., Yano, J. M., Fung, T. C. & Hsiao, E. Y. The microbiome and host behavior. *Annu. Rev. Neurosci.* 40, 21–49 (2017).
- Tierney, B. T. et al. The landscape of genetic content in the gut and oral human microbiome. *Cell Host Microbe* 26, 283–295.e8 (2019).
- Meganathan, R. & Kwon, O. Biosynthesis of menaquinone (vitamin K2) and ubiquinone (coenzyme Q). *EcoSal Plus* https://doi.org/10.1128/ecosalplus. 3.6.3.3 (2009).
- Hanke, M. L. & Kielian, T. Toll-like receptors in health and disease in the brain: mechanisms and therapeutic potential. *Clin. Sci.* **121**, 367–387 (2011).
- Sellge, G. & Kufer, T. A. PRR-signaling pathways: learning from microbial tactics. *Semin. Immunol.* 27, 75–84 (2015).
 Skaper, S. D., Facci, L., Zusso, M. & Giusti, P.
- Skaper, S. D., Facci, L., Zusso, M. & Giusti, P. An inflammation-centric view of neurological disease: beyond the neuron. *Front. Cell Neurosci.* 12, 72 (2018).
- Arentsen, T. et al. The bacterial peptidoglycan-sensing molecule Pglyrp2 modulates brain development and behavior. *Mol. Psychiatry* 22, 257–266 (2017).
- Vargas-Caraveo, A. et al. Lipopolysaccharide enters the rat brain by a lipoprotein-mediated transport mechanism in physiological conditions. *Sci. Rep.* 7, 13113 (2017).
- Bassi, G. S. et al. Lipopolysaccharide-induced sickness behaviour evaluated in different models of anxiety and innate fear in rats. *Basic Clin. Pharmacol. Toxicol.* 110, 359–369 (2012).
- Zhao, J. et al. Neuroinflammation induced by lipopolysaccharide causes cognitive impairment in mice. *Sci. Rep.* 9, 5790 (2019).
- O'Connor, J. C. et al. Lipopolysaccharide-induced depressive-like behavior is mediated by indoleamine 2,3-dioxygenase activation in mice. *Mol. Psychiatry* 14, 511–522 (2009).
- Romero, E. et al. Neurobehavioral and immunological consequences of prenatal immune activation in rats. Influence of antipsychotics. *Neuropsychopharmacology* **32**, 1791–1804 (2007).
- Izvolskaia, M., Sharova, V. & Zakharova, L. Prenatal programming of neuroendocrine system development by lipopolysaccharide: long-term effects. *Int. J. Mol. Sci.* 19, 3695 (2018).
- Caputi, V. & Giron, M. C. Microbiome–gut–brain axis and Toll-like receptors in Parkinson's disease. *Int. J. Mol. Sci.* **19**, 1689 (2018).
- Ohgi, Y., Futamura, T., Kikuchi, T. & Hashimoto, K. Effects of antidepressants on alternations in serum cytokines and depressive-like behavior in mice after lipopolysaccharide administration. *Pharmacol. Biochem. Behav.* 103, 853–859 (2013).

- Holzer, P. et al. Visceral inflammation and immune activation stress the brain. *Front. Immunol.* 8, 1613 (2017).
- Popoff, M. R. & Poulain, B. Bacterial toxins and the nervous system: neurotoxins and multipotential toxins interacting with neuronal cells. *Toxins* 2, 683–737 (2010).
- Kiu, R. & Hall, L. J. An update on the human and animal enteric pathogen *Clostridium perfringens*. *Emerg. Microbes. Infect.* https://doi.org/10.1038/ s41426-018-0144-8 (2018).
- Miyamoto, O. et al. *Clostridium perfringens* epsilon toxin causes excessive release of glutamate in the mouse hippocampus. *FEMS Microbiol. Lett.* 189, 109–113 (2000).
- Yang, N. J. & Chiu, I. M. Bacterial signaling to the nervous system via toxins and metabolites. *J. Mol. Biol.* 429, 587–605 (2017).
- Nagahama, M. & Sakurai, J. Distribution of labeled *Clostridium perfringens* epsilon toxin in mice. *Toxicon* 29, 211–217 (1991).
- Agata, N., Ohta, M., Mori, M. & Isobe, M. A novel dodecadepsipeptide, cereulide, is an emetic toxin of *Bacillus cereus. FEMS Microbiol. Lett.* **129**, 17–19 (1995).
- Sugiyama, H. & Hayama, T. Abdominal viscera as site of emetic action for staphylococcal enterotoxin in the monkey. J. Infect. Dis. 115, 330–336 (1965).
- Hu, D.-L. et al. Staphylococcal enterotoxin induces emesis through increasing serotonin release in intestine and it is downregulated by cannabinoid receptor 1. *Cell. Microbiol.* 9, 2267–2277 (2007).
- Friedland, R. P. & Chapman, M. R. The role of microbial amyloid in neurodegeneration. *PLoS Pathog.* 13, e1006654 (2017).
- Chapman, M. R. et al. Role of *Escherichia coli* curli operons in directing amyloid fiber formation. *Science* 295, 851–855 (2002).
- Collinson, S. K., Emödý, L., Müller, K. H., Trust, T. J. & Kay, W. W. Purification and characterization of thin, aggregative fimbriae from *Salmonella enteritidis*. *J. Bacteriol.* **173**, 4773–4781 (1991).
- Chen, S. G. et al. Exposure to the functional bacterial amyloid protein curli enhances α-synuclein aggregation in aged Fischer 344 rats and *Caenorhabditis elegans*. *Sci. Rep.* 6, 34477 (2016).
- Sci. Rep. 6, 34477 (2016).
 94. Russell, D. W. The enzymes, regulation, and genetics of bile acid synthesis. Annu. Rev. Biochem. 72, 137–174 (2003).
- 95. Hylemon, P. B. et al. Bile acids as regulatory molecules *J. Linid Res.* **50**, 1509–1520 (2009)
- molecules. J. Lipid Res. 50, 1509–1520 (2009).
 Keitel, V. et al. The bile acid receptor TGR5 (Cpbar-1) acts as a neurosteroid receptor in brain. *Glia* 58, 1794–1805 (2010).
- Yang, A. H., Ishii, I. & Chun, J. In vivo roles of lysophospholipid receptors revealed by gene targeting studies in mice. *Biochim. Biophys. Acta* 1582, 197–203 (2002).
- Mertens, K. L., Kalsbeek, A., Soeters, M. R. & Eggink, H. M. Bile acid signaling pathways from the enterohepatic circulation to the central nervous protection for the contral nervous
- system. Front. Neurosci. 11, 617 (2017).
 Singh, J., Metrani, R., Shivanagoudra, S. R., Jayaprakasha, G. K. & Patil, B. S. Review on bile acids: effects of the gut microbiome, interactions with dietary fiber, and alterations in the bioaccessibility of bioactive compounds. J. Agric. Food Chem. 67, 9124–9138 (2019).
- Philipp, B. Bacterial degradation of bile salts. Appl. Microbiol. Biotechnol. 89, 903–915 (2011).
- Lund, E. G., Guileyardo, J. M. & Russell, D. W. cDNA cloning of cholesterol 24-hydroxylase, a mediator of cholesterol homeostasis in the brain. *Proc. Natl Acad. Sci. USA* 96, 7238–7243 (1999).
- 102. Kim, H. J. et al. Common CYP7A1 promoter polymorphism associated with risk of neuromyelitis optica. *Neurobiol. Dis.* **37**, 349–355 (2010).
- Båvner, A. et al. On the mechanism of accumulation of cholestanol in the brain of mice with a disruption of sterol 27-hydroxylase. *J. Lipid Res.* 51, 2722–2730 (2010).
- 104. Kotti, T. J., Ramirez, D. M. O., Pfeiffer, B. E., Huber, K. M. & Russell, D. W. Brain cholesterol turnover required for geranylgeraniol production and learning in mice. *Proc. Natl Acad. Sci. USA* **103**, 3869–3874 (2006).
- McMillin, M. & DeMorrow, S. Effects of bile acids on neurological function and disease. *FASEB J.* 30, 3658–3668 (2016).

- McMillin, M. et al. TGR5 signaling reduces neuroinflammation during hepatic encephalopathy. *J. Neurochem.* 135, 565–576 (2015).
- MacLennan, A. J. et al. An essential role for the H218/ACR16/Edg-5/LPB2 sphingosine 1-phosphate receptor in neuronal excitability. *Eur. J. Neurosci.* 14, 203–209 (2001).
- 203–209 (2001).
 108. Gohlke, H., Schmitz, B., Sommerfeld, A., Reinehr, R. & Häussinger, D. a5β1-Integrins are sensors for tauroursodeoxycholic acid in hepatocytes. *Hepatology* 57, 1117–1129 (2013).
- 57, 1117–1129 (2013).
 109. Schubring, S. R., Fleischer, W., Lin, J. S., Haas, H. L. & Sergeeva, O. A. The bile steroid chenodeoxycholate is a potent antagonist at NMDA and CABAA receptors. *Neurosci. Lett.* 506, 322–326 (2012).
- Yanovski, Y. et al. Waking action of ursodeoxycholic acid (UDCA) involves histamine and GABAA receptor block. *PLoS ONE* 7, e42512 (2012).
 MNeilly, A. D. et al. Bile acids modulate glucocorticoid
- MNeilly, A. D. et al. Bile acids modulate glucocorticoid metabolism and the hypothalamic–pituitary–adrenal axis in obstructive jaundice. *J. Hepatol.* 52, 705–711 (2010).
- 112. Quinn, M. et al. Suppression of the HPA axis during extrahepatic biliary obstruction induces cholangiocyte proliferation in the rat. Am. J. Physiol. Gastrointest. Liver Physiol. **302**, C182–C193 (2011).
- 113. Hertel, J. et al. Integrated analyses of microbiome and longitudinal metabolome data reveal microbial–host interactions on sulfur metabolism in Parkinson's disease. *Cell Rep.* 29, 1767–1777.e8 (2019).
- 114. Nho, K. et al. Altered bile acid profile in mild cognitive impairment and Alzheimer's disease: relationship to neuroimaging and CSF biomarkers. *Alzheimers Dement.* 15, 232–244 (2019).
- 115. MahmoudianDehkordi, S. et al. Altered bile acid profile associates with cognitive impairment in Alzheimer's disease — an emerging role for gut microbiome. Alzheimers Dement. **15**, 76–92 (2019).
- 116. Ho, P. P. & Steinman, L. Obeticholic acid, a synthetic bile acid agonist of the farnesoid X receptor, attenuates experimental autoimmune encephalomyelitis. *Proc. Natl Acad. Sci. USA* **113**, 1600–1605 (2016).
- Bhargava, P. et al. Bile acid metabolism is altered in multiple sclerosis and supplementation ameliorates neuroinflammation. J. Clin. Invest. 130, 3467–3482 (2020).
- Wang, G. et al. Gut microbiota and relevant metabolites analysis in alcohol dependent mice. *Front. Microbiol.* 9, 1874 (2018).
- 119. Golubeva, A. V. et al. Microbiota-related changes in bile acid & tryptophan metabolism are associated with gastrointestinal dysfunction in a mouse model of autism. *EBioMedicine* 24, 166–178 (2017).
- 120. Swain, M. G., Patchev, V., Vergalla, J., Chrousos, G. & Jones, E. A. Suppression of hypothalamic–pituitary– adrenal axis responsiveness to stress in a rat model of acute cholestasis. *J. Clin. Invest.* **91**, 1903–1908 (1993).
- Rodrigues, C. M. P. et al. Neuroprotection by a bile acid in an acute stroke model in the rat. J. Cereb. Blood Flow. Metab. 22, 463–471 (2002).
- 122. Vaz, A. R. et al. Glycoursodeoxycholic acid reduces matrix metalloproteinase-9 and caspase-9 activation in a cellular model of superoxide dismutase-1 neurodegeneration. *Mol. Neurobiol.* **51**, 864–877 (2015).
- 123. Chakrabarti, A. et al. Transcriptomics driven lipidomics (TDL) identifies the microbiome-regulated targets of ileal lipid metabolism. *NPJ Syst. Biol. Appl.* **3**, 33 (2017).
- 124. Ghazalpour, A., Cespedes, I., Bennett, B. J. & Allayee, H. Expanding role of gut microbiota in lipid metabolism. *Curr. Opin. Lipidol.* 27, 141–147 (2016).
- Velagapudi, V. R. et al. The gut microbiota modulates host energy and lipid metabolism in mice. *J. Lipid Res.* 51, 1101–1112 (2010).
- 126. Lukovac, S. et al. Differential modulation by Akkermansia muciniphila and Faecalibacterium prausnitzii of host peripheral lipid metabolism and histone acetylation in mouse gut organoids. *mBio* 5, e01438–14 (2014).
- 127. Fu, J. et al. The gut microbiome contributes to a substantial proportion of the variation in blood lipids *Circ.Res.* **117**, 817–824 (2015).
- 128. An, D. et al. Sphingolipids from a symbiotic microbe regulate homeostasis of host intestinal natural killer T cells. *Cell* **156**, 123–133 (2014).
- 129. Lee, G., Hasan, M., Kwon, O.-S. & Jung, B. H. Identification of altered metabolic pathways during

disease progression in EAE mice via metabolomics and lipidomics. *Neuroscience* **416**, 74–87 (2019).

- Yano, J. M. et al. Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis. *Cell* 161, 264–276 (2015).
- Bean, L. A., Ianov, L. & Foster, T. C. Estrogen receptors, the hippocampus, and memory. *Neuroscientist* 20, 534–545 (2014).
- Nguyen, T.-V. Developmental effects of androgens in the human brain. J. Neuroendocrinol. https://doi.org/ 10.1111/jne.12486 (2018).
- 133. Oliveira, G. A. & Oliveira, R. F. Androgen modulation of social decision-making mechanisms in the brain: an integrative and embodied perspective. *Front. Neurosci.* 8, 29 (2014).
- 134. Bollinger, J. L., Salinas, I., Fender, E., Sengelaub, D. R. & Wellman, C. L. Gonadal hormones differentially regulate sex-specific stress effects on glia in the medial prefrontal cortex. *J. Neuroendocrinol.* https://doi.org/ 10.1111/jne.12762 (2019).
- 135. Cheng, J. et al. Exposure of hyperandrogen during pregnancy causes depression- and anxiety-like behaviors, and reduced hippocampal neurogenesis in rat offspring. *Front. Neurosci.* **13**, 436 (2019).
- Nead, K. T. Androgens and depression: a review and update. *Curr. Opin. Endocrinol. Diabetes Obes.* 26, 175–179 (2019).
- 137. Diotel, N. et al. Steroid transport, local synthesis, and signaling within the brain: roles in neurogenesis, neuroprotection, and sexual behaviors. *Front. Neurosci.* **12**, 84 (2018).
- 138. Zhu, B. T. & Conney, A. H. Functional role of estrogen metabolism in target cells: review and perspectives. *Carcinogenesis* 19, 1–27 (1998).
- Hellman, L. et al. The fate of hydrocortisone-4-C¹⁴ in man. J. Clin. Invest. 33, 1106–1115 (1954).
 Bokkenheuser, V. D. & Winter, J. Biotransformation of
- Bokkenheuser, V. D. & Winter, J. Biotransformation of steroid hormones by gut bacteria. *Am. J. Clin. Nutr.* 33, 2502–2506 (1980).
- Groh, H., Schade, K. & Hörhold-Schubert, C. Steroid metabolism with intestinal microorganisms. J. Basic Microbiol. 33, 59–72 (1993).
 Carcia-Gomez, E., González-Pedrajo, B. &
- Garcia-Gomez, E., Conzalez-Pedrajo, B. & Camacho-Arroyo, I. Role of sex steroid hormones in bacterial–host interactions. *BioMed Res. Internat.* https://doi.org/10.1155/2013/928290 (2013).
 Gloux, K. et al. A metagenomic β-glucuronidase
- 143. Gloux, K. et al. A metagenomic β-glucuronidase uncovers a core adaptive function of the human intestinal microbiome. *Proc. Natl Acad. Sci. USA* **108**, 4539–4546 (2011).
- 144. Dabek, M., McCrae, S. I., Stevens, V. J., Duncan, S. H. & Louis, P. Distribution of β-glucosidase and β-glucuronidase activity and of β-glucuronidase gene gus in human colonic bacteria. *FEMS Microbiol. Ecol.* **66**, 487–495 (2008).
- 145. Beaud, D., Tailliez, P. & Anba-Mondoloni, J. Genetic characterization of the β-glucuronidase enzyme from a human intestinal bacterium, *Ruminococcus gnavus*. *Microbiology* **151**, 2323–2330 (2005).
- 146. McIntosh, F. M. et al. Phylogenetic distribution of genes encoding β-glucuronidase activity in human colonic bacteria and the impact of diet on faecal glycosidase activities. *Environ. Microbiol.* 14, 1876–1887 (2012).
- 147. Ridlon, J. M. èt al. *Clostridium scindens*: a human gut microbe with a high potential to convert glucocorticoids into androgens. *J. Lipid Res.* 54, 2437–2449 (2013).
- Devendran, S., Mythen, S. M. & Ridlon, J. M. The desA and desB genes from Clostridium scindens ATCC 35704 encode steroid-17,20-desmolase. J. Lipid Res. 59, 1005–1014 (2018).
- 149. Winter, J. & Bokkenheuser, V. D. 21-Dehydroxylation of corticoids by anaerobic bacteria isolated from human fecal flora. *J. Steroid Biochem.* 9, 379–384 (1978).
- 150. Cerone-McLernon, A. M., Winter, J., Mosbach, E. H. & Bokkenheuser, V. D. Side-chain cleavage of cortisol by fecal flora. *Biochim. Biophys. Acta Lipids Lipid Metab.* 666, 341–347 (1981).
- 151. Ojanotko-Harri, A., Nikkari, T., Harrl, M.-P. & Paunio, K. Metabolism of progesterone and testosterone by *Bacillus cereus* strain Socransky 67 and *Streptococcus mutans* strain Ingbritt. Oral. Microbiol. Immunol. 5, 237–239 (1990).
- 152. Soory, M. Bacterial steroidogenesis by periodontal pathogens and the effect of bacterial enzymes on steroid conversions by human gingival fibroblasts in culture. *J. Periodontal Res.* **30**, 124–131 (1995).
- 153. Lombardi, P., Goldin, B., Boutin, E. & Gorbach, S. L. Metabolism of androgens and estrogens by human

fecal microorganisms. J. Steroid Biochem. 9, 795–801 (1978).

- 154. Järvenpää, P., Kosunen, T., Fotsis, T. & Adlercreutz, H. In vitro metabolism of estrogens by isolated intestinal micro-organisms and by human faecal microflora. *J. Steroid Biochem.* **13**, 345–349 (1980).
- 155. Flores, R. et al. Fecal microbial determinants of fecal and systemic estrogens and estrogen metabolites: a cross-sectional study. J. Transl Med. 10, 253 (2012).
- Plottel, C. S. & Blaser, M. J. Microbiome and malignancy. *Cell Host Microbe* 10, 324–335 (2011).
- 157. Fuhrman, B. J. et al. Associations of the fecal microbiome with urinary estrogens and estrogen metabolites in postmenopausal women. J. Clin. Endocrinol. Metab. **99**, 4632–4640 (2014).
- 158. Goedert, J. J. et al. Investigation of the association between the fecal microbiota and breast cancer in postmenopausal women: a population-based casecontrol pilot study. J. Natl Cancer Inst. https://doi.org/ 10.1093/jnci/djv147 (2015).
- Villa, A., Vegeto, E., Poletti, A. & Maggi, A. Estrogens, neuroinflammation, and neurodegeneration. *Endocr. Rev.* 37, 372–402 (2016).
- Baker, J. M., Al-Nakkash, L. & Herbst-Kralovetz, M. M. Estrogen-gut microbiome axis: physiological and clinical implications. *Maturitas* 103, 45–53 (2017).
 Kaliannan, K. et al. Estrogen-mediated gut
- 161. Kaliannan, K. et al. Estrogen-mediated gut microbiome alterations influence sexual dimorphism in metabolic syndrome in mice. *Microbiome* 6, 25 (2018).
- Markle, J. G. M. et al. Sex differences in the gut microbiome drive hormone-dependent regulation of autoimmunity. *Science* 339, 1084–1088 (2013).
- 163. Senghor, B., Sokhna, C., Ruimy, R. & Lagier, J.-C. Gut microbiota diversity according to dietary habits and geographical provenance. *Hum. Microbiome J.* **7–8**, 1–9 (2018).
- 164. Sasabe, J. et al. Interplay between microbial D-amino acids and host D-amino acid oxidase modifies murine mucosal defence and gut microbiota. *Nat. Microbiol.* 1, 16125 (2016).
- 165. Metges, C. C. Contribution of microbial amino acids to amino acid homeostasis of the host. J. Nutr. 130, 18575–1864S (2000).
- 166. Dodd, D. et al. A gut bacterial pathway metabolizes aromatic amino acids into nine circulating metabolites. *Nature* 551, 648–652 (2017).
- 167. Wikoff, W. R. et al. Metabolomics analysis reveals large effects of gut microflora on mammalian blood metabolites. PNAS 106, 3698–3703 (2009).
- 168. Asano, Y. et al. Critical role of gut microbiota in the production of biologically active, free catecholamines in the gut lumen of mice. *Am. J. Physiol. Gastrointest. Liver Physiol.* **303**, C1288–C1295 (2012).
- 169. Tsavkelova, E. A., Botvinko, I. V., Kudrin, V. S. & Oleskin, A. V. Detection of neurotransmitter amines in microorganisms with the use of high-performance liquid chromatography. *Dokl. Biochem.* **372**, 115–117 (2000).
- 170. Sperandio, V., Torres, A. G., Jarvis, B., Nataro, J. P. & Kaper, J. B. Bacteria–host communication: the language of hormones. *PNAS* **100**, 8951–8956 (2003).
- Kiraly, D. D. et al. Alterations of the host microbiome affect behavioral responses to cocaine. *Sci. Rep.* 6, 35455 (2016).
- 172. O'Farrell, K. & Harkin, A. Stress-related regulation of the kynurenine pathway: relevance to neuropsychiatric and degenerative disorders. *Neuropharmacology* **112**, 307–323 (2017).
- Jaglin, M. et al. Indole, a signaling molecule produced by the gut microbiota, negatively impacts emotional behaviors in rats. *Front. Neurosci.* **12**, 216 (2018).
 Zucchi, R., Chiellini, C., Scanlan, T. S. & Grandy, D. K.
- 174. Zucchi, R., Chiellini, G., Scanlan, T. S. & Grandy, D. K. Trace amine-associated receptors and their ligands. *Br. J. Pharmacol.* **149**, 967–978 (2006).
- 175. Schwarcz, R., Bruno, J. P., Muchowski, P. J. & Wu, H.-Q. Kynurenines in the mammalian brain: when physiology meets pathology. *Nat. Rev. Neurosci.* 13, 465–477 (2012).
- 176. Yanovsky, I. et al. Carbamate derivatives of indolines as cholinesterase inhibitors and antioxidants for the treatment of Alzheimer's disease. J. Med. Chem. 55, 10700–10715 (2012).
- 177. Adesso, S. et al. Indoxyl sulfate affects glial function increasing oxidative stress and neuroinflammation in chronic kidney disease: interaction between astrocytes and microglia. *Front. Pharmacol.* **8**, 370 (2017).
- Needham, B. D. et al. Plasma and fecal metabolite profiles in autism spectrum disorder. *bioRxiv* https://doi.org/10.1101/2020.05.17.098806 (2020).

- 179. Gabriele, S. et al. Urinary p-cresol is elevated in young French children with autism spectrum disorder a replication study. Biomarkers 19, 463–470 (2014).
- 180. Gacias, M. et al. Microbiota-driven transcriptional changes in prefrontal cortex override genetic
- differences in social behavior. eLife 5, e13442 (2016). 181. Zhu, L. et al. Structure and regulation of the gab gene cluster, involved in the γ -aminobutyric acid shunt, are controlled by a o54 factor in Bacillus thuringiensis J. Bacteriol. 192, 346–355 (2010).
- 182. O'Byrne, C. P. & Karatzas, K. A. G. The role of sigma B (gB) in the stress adaptations of Listeria monocytogenes: overlaps between stress adaptation and virulence. Adv. Appl. Microbiol. 65, 115-140 (2008).
- 183. Olson, C. A. et al. The gut microbiota mediates the anti-seizure effects of the ketogenic diet. Cell 173. 1728-1741 (2018).
- 184. Pokusaeva, K. et al. GABA-producing Bifidobacterium dentium modulates visceral sensitivity in the intestine. Neurogastroenterol. Motil. 29, e12904 (2017).
- 185. Strandwitz, P. et al. GABA-modulating bacteria of the human gut microbiota. Nat. Microbiol. 4, 396-403 (2019).
- 186. Horder, J. et al. Glutamate and GABA in autism spectrum disorder — a translational magnetic resonance spectroscopy study in man and rodent models. *Transl Psychiatry* 8, 106 (2018).
 187. Femenía, T., Gómez-Galán, M., Lindskog, M. &
- Magara, S. Dysfunctional hippocampal activity affects emotion and cognition in mood disorders. Brain Res.
- 1476, 58–70 (2012). 188. Soeiro-de-Souza, M. G. et al. Anterior cingulate glutamate-glutamine cycle metabolites are altered in euthymic bipolar I disorder. Eur. Neuropsychopharmacol. 25, 2221-2229 (2015).
- 189. Cherlyn, S. Y. T. et al. Genetic association studies of glutamate, GABA and related genes in schizophrenia and bipolar disorder: a decade of advance. Neurosci. Biobehav. Rev. 34, 958–977 (2010).
- 190. Williams, K., Zappia, A. M., Pritchett, D. B., Shen, Y. M. & Molinoff, P. B. Sensitivity of the N-methyl-Daspartate receptor to polyamines is controlled by NR2 subunits. *Mol. Pharmacol.* **45**, 803–809 (1994).
- 191. Matsumoto, M. et al. Impact of intestinal microbiota on intestinal luminal metabolome. Sci. Rep. 2, 233 (2012).
- 192. Sugiyama, Y. et al. Analysis of polyamine biosynthetic-and transport ability of human indigenous Bifidobacterium. Biosci. Biotechnol. Biochem. 82, 1606–1614 (2018).
- 193. Akasaka, N. & Fujiwara, S. The therapeutic and nutraceutical potential of agmatine, and its enhanced production using Aspergillus oryzae. Amino Acids https://doi.org/10.1007/s00726-019-02720-(2019).
- 194. Barua, S., Kim, J. Y., Kim, J. Y., Kim, J. H. & Lee, J. E. Therapeutic effect of agmatine on neurological disease: focus on ion channels and receptors Neurochem. Res. 44, 735–750 (2019)
- 195. Deka, G., Bharath, S. R., Savithri, H. S. & Murthy, M. R. N. Structural studies on the decameric *S. typhimurium* arginine decarboxylase (ADC): pyridoxal 5'-phosphate binding induces conformational changes. Biochem. Biophys. Res. Commun. 490, 1362-1368 (2017).
- 196. Andrell, J. et al. Crystal structure of the acid-induced arginine decarboxylase from Escherichia coli: reversible decamer assembly controls enzyme activity. Biochemistry 48, 3915–3927 (2009)
- 197. Wu, N., Su, R.-B. & Li, J. Agmatine and imidazoline receptors: their role in opioid analgesia, tolerance and dependence. *Cell Mol. Neurobiol.* **28**, 629–641 (2008). 198. Taksande, B. G. et al. Agmatine, an endogenous
- imidazoline receptor ligand modulates ethanol anxiolysis and withdrawal anxiety in rats. Eur. . Pharmacol. 637, 89–101 (2010).
- 199. Sameer, S., Chakraborty, S. & Ugale, R. Agmatine attenuates acquisition but not the expression of ethanol conditioned place preference in mice: a role for imidazoline receptors. Behav. Pharmacol. 24, 87-94 (2013).
- 200. Shopsin, B. The clinical antidepressant effect of exogenous agmatine is not reversed by parachlorophenylalanine: a pilot study. Acta Neuropsychiatr. 25, 113-118 (2013).
- 201. Gupta, V. K. et al. Restoring polyamines protects from age-induced memory impairment in an autophagy-dependent manner. Nat. Neurosci. 16, 1453–1460 (2013)
- 202. Kang, S. et al. Agmatine ameliorates type 2 diabetes induced-Alzheimer's disease-like alterations in high-fat

diet-fed mice via reactivation of blunted insulin

- signalling. *Neuropharmacology* **113**, 467–479 (2017). 203. Li, J., Doyle, K. M. & Tatlisumak, T. Polyamines in the brain: distribution, biological interactions, and their potential therapeutic role in brain ischaemia. Curr. Med. Chem. 14, 1807-1813 (2007).
- 204. Lyte, M. Microbial endocrinology: host-microbiota neuroendocrine interactions influencing brain and behavior. Gut Microbes 5, 381-389 (2014).
- 205. Koh, A., De Vadder, F., Kovatcheva-Datchary, P. & Bäckhed, F. From dietary fiber to host physiology: short-chain fatty acids as key bacterial metabolites. *Cell* **165**, 1332–1345 (2016).
- 206. Macfarlane, G. T. & Macfarlane, S. Bacteria, colonic fermentation, and gastrointestinal health. J. AOAC Int. 40, 50-60 (2019).
- 207. Sadler, R. et al. Short-chain fatty acids improve poststroke recovery via immunological mechanisms J. Neurosci. 40, 1162–1173 (2020).
- 208. Frost, G. et al. The short-chain fatty acid acetate reduces appetite via a central homeostatic mechanism. Nat. Commun. 5, 3611 (2014)
- 209. Wyss, M. T., Magistretti, P. J., Buck, A. & Weber, B. Labeled acetate as a marker of astrocytic metabolism. J. Cereb. Blood Flow. Metab. 31, 1668–1674 (2011).
- 210. Hoyles, L. et al. Microbiome-host systems interactions: protective effects of propionate upon the blood-brain barrier. Microbiome 6, 55 (2018) Duscha, A. et al. Propionic acid shapes the multiple 211
- sclerosis disease course by an immunomodulatory mechanism. Cell 180, 1067-1080 (2020).
- 212. MacFabe, D. F. et al. A novel rodent model of autism: intraventricular infusions of propionic acid increase locomotor activity and induce neuroinflammation and oxidative stress in discrete regions of adult rat brain. Am. J. Biochem. Biotechnol. 4, 146-166 (2008).
- 213. Shultz, S. R. et al. Intracerebroventricular injections of the enteric bacterial metabolic product propionic acid impair cognition and sensorimotor ability in the Long-Evans rat: further development of a rodent model of autism. Behav. Brain Res. 200, 33-41 (2009).
- 214. Sleiman, S. F. et al. Putting the 'HAT' back on survival signalling: the promises and challenges of HDAC inhibition in the treatment of neurological conditions. Expert Opin. Investigat. Drugs 18, 573-584 (2009).
- 215. Govindarajan, N., Agis-Balboa, R. C., Walter, J., Sananbenesi, F. & Fischer, A. Sodium butyrate improves memory function in an Alzheimer's disease mouse model when administered at an advanced stage of disease progression. J. Alzheimers Dis. 26, 187-197 (2011).
- 216. Kilgore, M. et al. Inhibitors of class 1 histore deacetylases reverse contextual memory deficits in a mouse model of Alzheimer's disease. Neuropsychopharmacology 35, 870-880 (2010).
- 217. da Silva, P. F. et al. Memory impairment induced by brain iron overload is accompanied by reduced H3K9 acetylation and ameliorated by sodium butyrate. *Neuroscience* **200**, 42–49 (2012).
- 218. Dash, P. K., Orsi, S. A. & Moore, A. N. Histone deactylase inhibition combined with behavioral therapy enhances learning and memory following traumatic brain injury. Neuroscience 163, 1-8 (2009).
- 219. Steckert, A. V. et al. Effects of sodium butyrate on aversive memory in rats submitted to sepsis. Neurosci. Lett. 595, 134-138 (2015).
- 220. Barichello, T. et al. Sodium butyrate prevents memory impairment by re-establishing BDNF and GDNF expression in experimental pneumococcal meningitis. Mol. Neurobiol. 52, 734-740 (2015).
- 221. Kim, H. J. & Chuang, D.-M. HDAC inhibitors mitigate ischemia-induced oligodendrocyte damage: potential roles of oligodendrogenesis, VEGF, and antiinflammation. Am. J. Transl Res. 6, 206-223 (2014).
- 222. Gardian, G. et al. Neuroprotective effects of phenylbutyrate in the N171-82Q transgenic mouse model of Huntington's disease. J. Biol. Chem. 280, 556-563 (2005)
- 223. Ferrante, R. J. et al. Histone deacetylase inhibition by sodium butyrate chemotherapy ameliorates the neurodegenerative phenotype in Huntington's disease
- mice. J. Neurosci. 23, 9418–9427 (2003).
 224. Kidd, S. K. & Schneider, J. S. Protection of dopaminergic cells from MPP⁺-mediated toxicity by histone deacetylase inhibition. Brain Res. 1354 172-178 (2010)
- 225. Crozier, A., Clifford, M. N. & Ashihara, H. Plant Secondary Metabolites: Occurrence, Structure and Role in the Human Diet (Wiley, 2008).
- 226. Manach, C., Williamson, G., Morand, C., Scalbert, A. & Rémésy, C. Bioavailability and bioefficacy of

polyphenols in humans. I. Review of 97 bioavailability studies. Am. J. Clin. Nutr. 81, 230S-242S (2005).

- 227. Sadeghi Ekbatan, S. et al. Absorption and metabolism of phenolics from digests of polyphenol-rich potato extracts using the Caco-2/HepG2 co-culture system Foods 7, 8 (2018).
- 228. Marín, L., Miguélez, E. M., Villar, C. J. & Lombó, F. Bioavailability of dietary polyphenols and gut microbiota metabolism: antimicrobial properties Biomed. Res. Int. https://doi.org/10.1155/2015/ 905215 (2015). 229. Liu, Z. & Hu, M. Natural polyphenol disposition via
- coupled metabolic pathways. Expert Opin. Drug Metab. Toxicol. 3, 389-406 (2007).
- 230. Ferruzzi, M. G. et al. Bioavailability of gallic acid and catechins from grape seed polyphenol extract is improved by repeated dosing in rats: implications for treatment in Alzheimer's disease. J. Alzheimers Dis. 18, 113-124 (2009).
- 231. Ho, L. et al. Identification of brain-targeted bioactive dietary quercetin-3-O-glucuronide as a novel intervention for Alzheimer's disease. FASEB J. 27, 769-781 (2013)
- 232. Wang, J. et al. Brain-targeted proanthocyanidin metabolites for Alzheimer's disease treatment. J. Neurosci. 32, 5144–5150 (2012).
- 233. Wang, D. et al. Role of intestinal microbiota in the generation of polyphenol-derived phenolic acid mediated attenuation of Alzheimer's disease β-amyloid oligomerization. Mol. Nutr. Food Res. 59, 1025-1040 (2015)
- 234. Loureiro, J. A. et al. Resveratrol and grape extractloaded solid lipid nanoparticles for the treatment of Alzheimer's disease. *Molecules* **22**, 277 (2017).
- 235. Wang, J. et al. Targeting multiple pathogenic mechanisms with polyphenols for the treatment of Alzheimer's disease - experimental approach and therapeutic implications. Front. Aging Neurosci. 6, 42 (2014)
- 236. Wang, J. et al. Epigenetic modulation of inflammation and synaptic plasticity promotes resilience against stress in mice. Nat. Commun. 9, 477 (2018).
- 237. Tomaro-Duchesneau, C. et al. Probiotic ferulic acid esterase active Lactobacillus fermentum NCIMB 5221 APA microcapsules for oral delivery: preparation and in vitro characterization. Pharmaceuticals 5 236-248 (2012).
- 238. Ren, Z. et al. Ferulic acid exerts neuroprotective effects against cerebral ischemia/reperfusion-induced injury via antioxidant and anti-apoptotic mechanisms in vitro and in vivo. Int. J. Mol. Med. 40, 1444-1456 (2017)
- 239. Mori, T., Koyama, N., Guillot-Sestier, M.-V., Tan, J. δ Town, T. Ferulic acid is a nutraceutical β-secretase modulator that improves behavioral impairment and Alzheimer-like pathology in transgenic mice. PLoS ONE 8, e55774 (2013).
- 240. Zeni, A. L. B., Camargo, A. & Dalmagro, A. P. Ferulic acid reverses depression-like behavior and oxidative stress induced by chronic corticosterone treatment in mice. Steroids 125, 131-136 (2017).
- 241. Wang, J. et al. Cocoa extracts reduce oligomerization of amyloid- β : implications for cognitive improvement in Alzheimer's disease. J. Alzheimers Dis. **41**, 643–650 (2014).
- 242. Santa-Maria, I. et al. GSPE interferes with tau aggregation in vivo: implication for treating tauopathy. Neurobiol. Aging 33, 2072-2081 (2012).
- 243. Bode, L. M. et al. In vivo and in vitro metabolism of trans-resveratrol by human gut microbiota. Am. J. Clin. Nutr. 97, 295–309 (2013).
- 244. Zhang, L.-F. et al. Resveratrol alleviates motor and cognitive deficits and neuropathology in the A53T α-synuclein mouse model of Parkinson's disease. Food Funct. 9, 6414–6426 (2018).
- 245. Sampson, T. R. et al. A gut bacterial amyloid promotes $\alpha\mbox{-synuclein}$ aggregation and motor impairment in mice. eLife 9, e53111 (2020).
- 246, Clavel, T., Borrmann, D., Braune, A., Doré, J. & Blaut, M. Occurrence and activity of human intestinal bacteria involved in the conversion of dietary lignans. Anaerobe 12, 140-147 (2006).
- 247. Rafii, F. The role of colonic bacteria in the metabolism of the natural isoflavone daidzin to equol. Metabolites 5, 56-73 (2015).
- 248. Rietjens, I. M. C. M., Louisse, J. & Beekmann, K. The potential health effects of dietary phytoestrogens.
- Br. J. Pharmacol. 174, 1263–1280 (2017).
 249. Sakai, T. & Kogiso, M. Soy isoflavones and immunity. J. Med. Invest. 55, 167–173 (2008).
- 250. Mueller, S. O., Simon, S., Chae, K., Metzler, M. & Korach, K. S. Phytoestrogens and their human

PERSPECTIVES

metabolites show distinct agonistic and antagonistic properties on estrogen receptor alpha (ERa) and ER β in human cells. *Toxicol. Sci.* **80**, 14–25 (2004).

- Cooke, P. S., Selvaraj, V. & Yellayi, S. Genistein, estrogen receptors, and the acquired immune response. J. Nutr. 136, 704–708 (2006).
- response. J. Nutr. **136**, 704–708 (2006). 252. Little, M. S., Pellock, S. J., Walton, W. G., Tripathy, A. & Redinbo, M. R. Structural basis for the regulation of β -glucuronidase expression by human gut Enterobacteriaceae. *Proc. Natl Acad. Sci. USA* **115**, E152–E161 (2018).
- E152–E161 (2018). 253. Roberts, M. S., Magnusson, B. M., Burczynski, F. J. & Weiss, M. Enterohepatic circulation. *Clin. Pharmacokinet.* **41**, 751–790 (2002).
- 254. Krishnaswamy, S. et al. Serotonin (5-hydroxytryptamine) glucuronidation in vitro: assay development, human liver microsome activities and species differences. *Xenobiotica* 33, 169–180 (2003).
- 255. Guthrie, L., Wolfson, S. & Kelly, L. The human gut chemical landscape predicts microbe-mediated biotransformation of foods and drugs. *eLife* 8, e42866 (2019).
- 256. Winter, J. & Bokkenheuser, V. D. Bacterial metabolism of natural and synthetic sex hormones undergoing enterohepatic circulation. *J. Steroid Biochem.* 27, 1145–1149 (1987).
- 257. Magnúsdóttir, S., Ravcheev, D., de Crécy-Lagard, V. & Thiele, I. Systematic genome assessment of B-vitamin biosynthesis suggests co-operation among gut microbes. *Front. Genet.* **6**, 148 (2015).
- 258. Rowland, I. et al. Gut microbiota functions: metabolism of nutrients and other food components. *Eur. J. Nutr.* **57**, 1–24 (2018).
- LeBlanc, J. G. et al. Bacteria as vitamin suppliers to their host: a gut microbiota perspective. *Curr. Opin. Biotechnol.* 24, 160–168 (2013).
 Hiratsuka, T. et al. An alternative menaguinone
- Hiratsuka, T. et al. An alternative menaquinone biosynthetic pathway operating in microorganisms. *Science* 321, 1670–1673 (2008).
 Ferland, G. Vitamin K and brain function.
- Semin. Thromb. Hemost. **39**, 849–855 (2013).
 Derrien, M. et al. Modulation of mucosal immune response, tolerance, and proliferation in mice colonized by the mucin-degrader Akkermansia
- muciniphila. Front. Microbiol. 2, 166 (2011).
 263. Yang, S., Minkler, P. & Hoppel, C. *cis*-3,4-Methyleneheptanoylcarnitine: characterization and verification of the C8:1 acylcarnitine in human urine. *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* 857, 251–258 (2007).
- 264. Zhang, L. S. & Davies, S. S. Microbial metabolism of dietary components to bioactive metabolites: opportunities for new therapeutic interventions. *Genome Med.* **8**, 46 (2016).
- Ogawa, J. et al. Production of conjugated fatty acids by lactic acid bacteria. J. Biosci. Bioeng. 100, 355–364 (2005).
- 266. Sberro, H. et al. Large-scale analyses of human microbiomes reveal thousands of small, novel genes. *Cell* **178**, 1245–1259.e14 (2019).
- Bercik, P. et al. The anxiolytic effect of *Bifidobacterium longum* NCC3001 involves vagal pathways for gut– brain communication. *Neurogastroenterol. Motil.* 23, 1132–1139 (2011).
- 268. Sgritta, M. et al. Mechanisms underlying microbialmediated changes in social behavior in mouse models of autism spectrum disorder. *Neuron* **101**, 246–259. e6 (2019).
- Goehler, L. E. et al. Activation in vagal afferents and central autonomic pathways: early responses to intestinal infection with *Campylobacter jejuni*. *Brain Behav. Immun.* 19, 334–344 (2005).
- Gershon, M. D. & Tack, J. The serotonin signaling system: from basic understanding to drug development for functional Gl disorders. *Gastroenterology* 132, 397–414 (2007).
- 271. Vadder, F. D. et al. Gut microbiota regulates maturation of the adult enteric nervous system via enteric serotonin networks. *Proc. Natl Acad. Sci. USA* 115, 6458–6463 (2018).
- Sjögren, K. et al. The gut microbiota regulates bone mass in mice. J. Bone Miner. Res. 27, 1357–1367 (2012).
- 273. Tian, P., Wang, G., Zhao, J., Zhang, H. & Chen, W. Bifidobacterium with the role of 5-hydroxytryptophan synthesis regulation alleviates the symptom of depression and related microbiota dysbiosis. J. Nutr. Biochem. 66, 43–51 (2019).
- 274. Lund, M. L. et al. Enterochromaffin 5-HT cells a major target for GLP-1 and gut microbial metabolites. *Mol. Metab.* **11**, 70–83 (2018).

- 275. Wang, H. et al. TLR2 plays a pivotal role in mediating mucosal serotonin production in the gut. *J. Immunol.* **202**, 3041–3052 (2019).
- Kidd, M. et al. Luminal regulation of normal and neoplastic human EC cell serotonin release is mediated by bile salts, amines, tastants, and olfactants. *Am. J. Physiol. Gastrointest. Liver Physiol.* 295, G260–G272 (2008).
- 277. Tsuruta, T. et al. Organoids as an ex vivo model for studying the serotonin system in the murine small intestine and colon epithelium. *Biochem. Biophys. Res. Commun.* **474**, 161–167 (2016).
- Reigstad, C. S. et al. Gut microbes promote colonic serotonin production through an effect of short-chain fatty acids on enterochromaffin cells. *FASEB J.* 29, 1395–1403 (2015).
- Neufeld, K.-A. M. et al. Oral selective serotonin reuptake inhibitors activate vagus nerve dependent gut-brain signalling. *Sci. Rep.* 9, 1–11 (2019).
- Ma, Q. et al. Impact of microbiota on central nervous system and neurological diseases: the gut-brain axis. *J. Neuroinflammation* 16, 53 (2019).
- 281. Frenois, F. et al. Lipopolysaccharide induces delayed FosB/∆FosB immunostaining within the mouse extended amygdala, hippocampus and hypothalamus, that parallel the expression of depressive-like behavior. *Psychoneuroendocrinology* **32**, 516–531 (2007).
- 282. Macfarlane, S., Cleary, S., Bahrami, B., Reynolds, N. & Macfarlane, G. T. Synbiotic consumption changes the metabolism and composition of the gut microbiota in older people and modifies inflammatory processes: a randomised, double-blind, placebo-controlled crossover study. *Aliment. Pharmacol. Ther.* **38**, 804–816 (2013).
- Kuo, S.-M. The interplay between fiber and the intestinal microbiome in the inflammatory response. *Adv. Nutr.* 4, 16–28 (2013).
- Adv. Nutr. 4, 16–28 (2013).
 284. Lee, S. U. et al. β-Arrestin 2 mediates G proteincoupled receptor 43 signals to nuclear factor-xB. *Biol. Pharm. Bull.* 36, 1754–1759 (2013).
- Patnala, R., Arumugam, T. V., Gupta, N. & Dheen, S. T. HDAC inhibitor sodium butyrate-mediated epigenetic regulation enhances neuroprotective function of microglia during ischemic stroke. *Mol. Neurobiol.* 54, 6391–6411 (2017).
- Erny, D. et al. Host microbiota constantly control maturation and function of microglia in the CNS. *Nat. Neurosci.* 18, 965–977 (2015).
- Borges, G., Lean, M. E. J., Roberts, S. A. & Crozier, A. Bioavailability of dietary (poly)phenols: a study with ileostomists to discriminate between absorption in small and large intestine. *Food Funct.* 4, 754–762 (2013).
- Pimpão, R. C., Ventura, M. R., Ferreira, R. B., Williamson, G. & Santos, C. N. Phenolic sulfates as new and highly abundant metabolites in human plasma after ingestion of a mixed berry fruit purée. *Br. J. Nutr.* **113**, 454–463 (2015).
- Gasperotti, M. et al. Fate of microbial metabolites of dietary polyphenols in rats: is the brain their target destination? ACS Chem. Neurosci. 6, 1341–1352 (2015).
- 290. Chen, T.-Y. et al. Plasma bioavailability and regional brain distribution of polyphenols from apple/grape seed and bilberry extracts in a young swine model. *Mol. Nutr. Food Res.* **59**, 2432–2447 (2015).
- Figueira, I. et al. Polyphenols journey through blood– brain barrier towards neuronal protection. *Sci. Rep.* 7, 1–16 (2017).
- 292. Youdim, K. A. et al. Interaction between flavonoids and the blood–brain barrier: in vitro studies. *J. Neurochem.* 85, 180–192 (2003).
- 293. Zhao, W. et al. Novel application of brain-targeting polyphenol compounds in sleep deprivation-induced cognitive dysfunction. *Neurochem. Int.* 89, 191–197 (2015).
- 294. Strandwitz, P. Neurotransmitter modulation by the gut microbiota. *Brain Res.* **1693**, 128–133 (2018).
- Cameron, J. S. et al. Toll-like receptor 3 is a potent negative regulator of axonal growth in mammals. *J. Neurosci.* 27, 13033–13041 (2007).
 Ma, Y. et al. Toll-like receptor 8 functions as a negative
- 296. Ma, Y. et al. Toll-like receptor 8 functions as a negative regulator of neurite outgrowth and inducer of neuronal apoptosis. J. Cell Biol. **175**, 209–215 (2006).
- Okun, E. et al. TLR2 activation inhibits embryonic neural progenitor cell proliferation. *J. Neurochem.* 114, 462–474 (2010).
- 298. Janssens, Y. et al. Screening of quorum sensing peptides for biological effects in neuronal cells. *Peptides* **101**, 150–156 (2018).

- 299. Rothhammer, V. et al. Type I interferons and microbial metabolites of tryptophan modulate astrocyte activity and central nervous system inflammation via the aryl hydrocarbon receptor. *Nat. Med.* **22**, 586–597 (2016).
- Rothhammer, V. et al. Microglial control of astrocytes in response to microbial metabolites. *Nature* 557, 724 (2018).
- 301. Liu, F., Horton-Sparks, K., Hull, V., Li, R. W. & Martínez-Cerdeño, V. The valproic acid rat model of autism presents with gut bacterial dysbiosis similar to that in human autism. *Mol. Autism* 9, 61 (2018).
- 302. Ho, M.-F. et al. Ketamine and ketamine metabolites as novel estrogen receptor ligands: induction of cytochrome P450 and AMPA glutamate receptor gene expression. *Biochem. Pharmacol.* **152**, 279–292 (2018).
- 303. Yang, T. et al. Butyrate regulates inflammatory cytokine expression without affecting oxidative respiration in primary astrocytes from spontaneously hypertensive rats. *Physiol. Rep.* 6, e13732 (2018).
- hypertensive rats. *Physiol. Rep.* **6**, e13732 (2018). 304. Xiang, Y. et al. Acetylpuerarin inhibits oxygen-glucose deprivation-induced neuroinflammation of rat primary astrocytes via the suppression of HIF-1 signaling. *Exp. Ther. Med.* **16**, 2689–2695 (2018).
- 305. Xiang, Y. et al. Anti-inflammatory effect of acetylpuerarin on eicosanoid signaling pathway in primary rat astrocytes. J. Mol. Neurosci. 52, 577–585 (2014).
- 306. Xin, Y. et al. Effects of oligosaccharides from *Morinda* officinalis on gut microbiota and metabolome of APP/ PS1 transgenic mice. *Front. Neurol.* 9, 412 (2018).
- 307. Chen, H. et al. Gut microbiota interventions with Clostridium butyricum and norfloxacin modulate immune response in experimental autoimmune encephalomyelitis mice. Front. Immunol. 10, 1662 (2019).
- Al-Ghezi, Z. Z., Busbee, P. B., Alghetaa, H., Nagarkatti, P. S. & Nagarkatti, M. Combination of cannabinoids, &-9-tetrahydrocannabinol (THC) and cannabidiol (CBD), mitigates experimental autoimmune encephalomyelitis (EAE) by altering the gut microbiome. *Brain Behav. Immun.* https://doi.org/ 10.1016/j.bbi.2019.07.028 (2019).
- 309. Gandy, K. A. O., Zhang, J., Nagarkatti, P. & Nagarkatti, M. The role of gut microbiota in shaping the relapse-remitting and chronic–progressive forms of multiple sclerosis in mouse models. *Sci. Rep.* 9, 1–17 (2019).
- Mangalam, A. et al. Human gut-derived commensal bacteria suppress CNS inflammatory and demyelinating disease. *Cell Rep.* 20, 1269–1277 (2017).
- Poisson, L. M. et al. Untargeted plasma metabolomics identifies endogenous metabolite with drug-like properties in chronic animal model of multiple sclerosis. *J. Biol. Chem.* **290**, 30697–30712 (2015).
- 313. Khalaj, A. J., Hasselmann, J., Augello, C., Moore, S. & Tiwari-Woodruff, S. K. Nudging oligodendrocyte intrinsic signaling to remyelinate and repair: estrogen receptor ligand effects. *J. Steroid Biochem. Mol. Biol.* 160, 43–52 (2016).
 314. Khalaj, A. J. et al. Estrogen receptor (ER) β expression
- 314. Khalaj, A. J. et al. Estrogen receptor (ER) β expression in oligodendrocytes is required for attenuation of clinical disease by an ERβ ligand. *PNAS* **110**, 19125–19130 (2013).
- 315. Takao, T. et al. 17β-Estradiol protects oligodendrocytes from cytotoxicity induced cell death. *J. Neurochem.* 89, 660–673 (2004).
- 316. Voskuhl, R. R. et al. Cene expression in oligodendrocytes during remyelination reveals cholesterol homeostasis as a therapeutic target in multiple sclerosis. *Proc. Natl Acad. Sci. USA* **116**, 10130–10139 (2019).
- Rankin, K. A. et al. Selective estrogen receptor modulators enhance CNS remyelination independent of estrogen receptors. *J. Neurosci.* **39**, 2184–2194 (2019).
- Abbott, N. J., Rönnbäck, L. & Hansson, E. Astrocyte– endothelial interactions at the blood–brain barrier. *Nat. Rev. Neurosci.* 7, 41–53 (2006).
- 319. Jin, L., Nation, R. L., Li, J. & Nicolazzo, J. A. Speciesdependent blood–brain barrier disruption of lipopolysaccharide: amelioration by colistin in vitro and in vivo. *Antimicrob. Agents Chemother.* 57, 4336–4342 (2013).

- 320. Braniste, V. et al. The gut microbiota influences blood– brain barrier permeability in mice. *Sci. Transl Med.* **6**, 263ra158 (2014).
- 321. Tang, A. T. et al. Endothelial TLR4 and the microbiome drive cerebral cavernous malformations. Nature 545,
- 305–310 (2017).
 322. Wolf, S. A., Boddeke, H. W. G. M. & Kettenmann, H. Microglia in physiology and disease. *Annu. Rev. Physiol.* **79**, 619–643 (2017).
- 323. Zhong, L.-M. et al. Resveratrol inhibits inflammatory responses via the mammalian target of rapamycin signaling pathway in cultured LPS-stimulated microglial cells. *PLoS ONE* **7**, e32195 (2012). 324. Catorce, M. N. & Gevorkian, G. LPS-induced murine
- neuroinflammation model: main features and

suitability for pre-clinical assessment of nutraceuticals. *Curr. Neuropharmacol.* **14**, 155–164 (2016).

(2016).
325. Yanguas-Casás, N., Barreda-Manso, M. A., Nieto-Sampedro, M. & Romero-Ramírez, L. TUDCA: an agonist of the bile acid receptor CPBAR1/TCR5 with anti-inflammatory effects in microglial cells. *J. Cell. Physiol.* 232, 2231–2245 (2017).

Author contributions

S.K.M. and B.D.N. researched data for the article and made substantial contributions to the discussion of content, writing, reviewing and editing of the manuscript before submission. R.K.-D. contributed to the review and editing of the manuscript before submission.

Competing interests

S.K.M. has financial interest in Axial Biotherapeutics. B.D.N. and R.K.-D. declare no competing interests.

Peer review information

Nature Reviews Neuroscience thanks Peter Holzer, who co-reviewed with Aitak Farzi; John Cryan; and Mauro Costa-Mattioli for their contribution to the peer review of this work.

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